Retinitis pigmentosa and stem cell therapy

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warrant further investigation.

trials, while also highlighting the existing limitations that

based on rigorous animal experimentation and clinical

substantiates the efficacy and safety of stem cell therapy

histopathological changes associated with RP, this study

to solve various problems faced by this approach in the

transplantation therapy, and further research is needed

cytopathia, which brings great challenges to stem cell

number of animal experiments and clinical trials based on

can be derived from stem cells. In recent years, a large

of RPE therapy since both RPE cells and photoreceptors

therapy, complication therapy, and stem cell therapy have been

INTRODUCTION

Retinitis pigmentosa (RP) is a group of hereditary retinal
disease characterized by damage to retinal photoreceptor

cells and retinal pigment epithelial (RPE) cells. Clinically, it is

characterized by night blindness, progressive reduced visual

field, pigmentary retinopathy, and poor photoreceptor function.
The disease usually occurs in both eyes, and in rare cases, it

occurs in one eye. The prevalence of RP ranges from 1/7000
to 1/3000 worldwide and about 1/4000 in China[1]. It usually
begins before the age of 30, most commonly in childhood or
adolescence, and becomes worse in adolescence until middle
age or old age due to macular involvement and severe visual
impairment and blindness. One of the most well-known
treatments for this disease is tissue therapy. However, with the
development of medicine, various therapeutic methods such as
gene therapy, retinal prosthesis, optogenetic therapy, drug
therapy, complication therapy, and stem cell therapy have been
proposed.

PATHOGENESIS OF RETINITIS PIGMENTOSA

A genetic abnormality is currently considered to be the main
cause of RP. Abnormal genes affect retinal photoreceptor

or RPE cells, weakening intracellular material transport,
molecular movement, and light conduction between

photoreceptor and RPE cells, ultimately leading to visual

impairment[2]. The genetic basis and mutations that cause RP

are complex and highly heterogeneous. More than 70 genes

have been identified as associated with RP, typically each

with a different disease-causing mutation[3-4]. According to the

latest findings, there are three types of autosomal recessive,
dominant, and X-linked recessive inheritance, with autosomal
recessive inheritance being the most common. Numerous

studies have shown that mutations in seven known splicing

factors are associated with autosomal dominant RP, including

pre-mRNA processing factor 3 (PRPF3), PRPF4, PRPF6,

PRPF8, PRPF31, small nuclear ribonucleoprotein particle 200

(SNRNP200), and pre-mRNA 9 (RP9; Table 1)[5-15]. Mutations
in PRPF31, are the most common among these, accounting for 6%-11.1% of autosomal dominant RP\cite{10}. PRPF31 is a highly conserved gene. Studies have found that homozygous PRPF31 knockout mice and zebrafish are embryonically lethal, indicating the importance of PRPF31. The present studies reveal that PRPF31-RP demonstrates incomplete exodominance due to haploinsufficiency, wherein the disease is attributed to reduced gene expression levels of the mutated allele\cite{9-12}. Xi et al\cite{12} found that carriers of the PRPF31 heterozygous mutation may be asymptomatic if the wild-type (WT) allele produces enough PRPF31 to maintain normal retinal function or may progress to blindness if the remaining PRPF31 level falls below a critical threshold.

### HISTOPATHOLOGICAL FEATURES

The most important changes in RP visible under the light microscope are the progressive degeneration of the upper cortex of the retinal nerve, especially the rod cells\cite{16}. Early studies showed that the lesions spread from the outer retinal layer inward, extending from the equator to the macula, eventually penetrating the nerve cell layer and leading to atrophy of the entire retina. In the early stage of the disease, the rods and pigment epithelial cells in the equatorial region, as well as the photoreceptor cells, simultaneously underwent degeneration and proliferation, and the proliferative epithelial cells and macrophages migrated to the vascular layer in the retina and near the veins. Degeneration and hyperplasia occur in the pigment epithelium, as the loss or accumulation of pigment that migrates to the inner retina\cite{17}. Proliferative epithelial cells, macrophages, and free pigment particles accumulated around the retinal blood vessels, and the outer membrane of the retinal blood vessels experienced hyaloid degeneration and thickening; the tube diameter was significantly reduced; and the lumen was narrow or even completely blocked. The choroidal blood vessels can then be sclerotized to varying degrees, and the late choroidal capillaries disappear completely or partially. Occlusion of the central artery of the optic nerve can result in optic neuropathy.

### Table 1 Causes of RP from different splicing factors

<table>
<thead>
<tr>
<th>Types of splicing factors</th>
<th>Causes of RP</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>PRPF3</td>
<td>Altered PRPF3 amino acids are highly conserved in all known PRPF3 orthologues</td>
<td>[5]</td>
</tr>
<tr>
<td>PRPF4</td>
<td>Haploinsufficiency and dominant-negative effects</td>
<td>[6]</td>
</tr>
<tr>
<td>PRPF6</td>
<td>Abnormal localization of endogenous PRPF6 within the nucleus</td>
<td>[7]</td>
</tr>
<tr>
<td>PRPF8</td>
<td>C-terminus exhibit a high degree of conservation</td>
<td>[8]</td>
</tr>
<tr>
<td>PRPF31</td>
<td>Incomplete penetrance; reduced levels of gene expression from the mutated allele</td>
<td>[9-12]</td>
</tr>
<tr>
<td>PR9</td>
<td>Proliferation and migration were decreased; FSCN2 and BBS2 were down-regulated in the mutated cells; pre-mRNA splicing of the FSCN2 gene was markedly affected</td>
<td>[13-14]</td>
</tr>
<tr>
<td>SNRNP200</td>
<td>A defect in hBrr2-dependent RNA unwinding and a consequent defect in spliceosome activation</td>
<td>[15]</td>
</tr>
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</table>

PRPF3: Pre-mRNA processing factor 3; PR9: Pre-mRNA 9; SNRNP200: Small nuclear ribonucleoprotein particle 200; FSCN2: Fascin actin-bunding protein 2; BBS2: Bardet-biedi syndrome 2; RP: Retinitis pigmentosa.
STEM CELL THERAPY OF RETINITIS PIGMENTOSA

Photoreceptor and Retinal Pigment Epithelium Cells  The retina is responsible for the conversion of light signals into electrical signals. Visual signals originate in photoreceptor cells, which mainly express photosensitive proteins in the outer segments. The human retina has two types of light-sensitive cells, rods and cones, which are responsible for dim vision and daylight vision (including color), respectively. The differentiation and proper functioning of photoreceptor cells and RPE cells are mutually dependent. During development, the neuroepithelial cells consist of two layers, which will give rise to the RPE cells and the neuronal retina. The absence of synaptic input and nutritional factors has been observed to inevitably result in transneuronal degeneration of retinal neurons due to photoreceptor death. However, even in cases of severe RP, approximately 30% of ganglion cells and around 80% of inner layer neurons remain unaffected. As a result, treatments for transplanting photoreceptors into the subretinal space and integrating them into the host retina by establishing synapses with the host bipolar cells began to emerge.

Stem Cell Therapy  Stem cells are pluripotent cells with the ability for targeted differentiation and proliferation. When the internal environment of the body changes, the differentiation potential of stem cells can be activated through various signaling pathways in the body and targeted differentiation into different cells and tissues, which is why stem cells are called universal cells. Because stem cells have the potential to replace damaged or missing retinal cells, they can be stimulated to differentiate into functional retinal photoreceptor cells and then injected into the retina to replace the damaged cells and repair the normal retinal tissue structure. Therefore, recent studies have shown that stem cell therapy is more commonly used and effective in RP because it can effectively promote retinal cell survival and inhibit the inflammatory response.

The emergence of pluripotent stem cells, from the initial embryonic stem cells (ESCs) to the human induced pluripotent stem cells (hiPSCs) currently being studied, provides a new idea for the treatment of RP. Numerous studies have shown that these cells can differentiate into several major retinal cell types, including photoreceptor cells. The emergence of pluripotent stem cells, initially ESCs and more recently hiPSCs, has provided a promising alternative source for attempting photoreceptor regeneration through cell transplantation. In addition, adult retinal pigment epithelial stem cells (RPESCs) offer a distinct lineage source for deriving RPE cells compared to ESCs and hiPSCs. Recent studies have demonstrated the presence of RPESCs in RPE cell cultures isolated from deceased human eyes. Under appropriate culture conditions, these stem cells can differentiate into functional RPE cells once again.

Singh et al. research suggests that adult RPESC-derived RPE cells may be a safer option than those derived from ESCs and hiPSCs; however, they may present other challenges as well. For instance, recent evidence indicates that cadaveric RPE cells may retain intracellular phenotypes associated with aging and disease, which could potentially limit their effectiveness as therapeutic substrates. The latest endeavor in this field aims to target the genetic form of RP by utilizing a specific type of cell derived from the eye, known as retinal progenitor cells (RPCs), which are isolated from fetal human eyes. In comparison to stem cells derived from bone marrow or neural stem cells, RPC transplantation may offer a more targeted treatment mechanism for retinal degeneration. Current clinical evidence suggests that transplanted RPCs retain the potential to differentiate into certain types of retinal cells, albeit with limited efficiency. Therefore, subretinal RPC transplantation has the potential to achieve permanent tissue repair through the actual regeneration of photoreceptor cells.

Animal Experiment  Animal studies have found that pluripotent stem cells, when used in mouse models of RP, have the potential not only to survive but also to differentiate into and function as photoreceptor cells. For example, the study conducted by Zhang et al. revealed a significant increase in the survival rate of photoreceptors and a notable enhancement in visual function following the intravitreal injection of mesystimal stem cell (MSCs) in mice. Liang et al. conducted animal experiments with umbilical cord MSCs (UCMSCs) and concluded that intravenous injection of UCMSCs could delay retinal degeneration and protect vision in rats. Brown et al. employed an innovative approach to investigate the therapeutic efficacy of original MSCs-derived RPCs in rd12 mouse models with retinal degeneration. Their findings demonstrated that transplanted RPCs effectively attenuated inflammation, provided retinal protection, and facilitated neurogenesis, ultimately leading to improvements in both retinal structure and physiological function in rd12 mice. Dezfuly et al. established a progressive acute retinal injury model by intravitreal injection of sodium iodate in rats to compare the therapeutic effects of human adipose-derived stem cells (hADSCs) and their secretome on in vivo models of sodium iodate-induced retinal neurodegeneration. The results demonstrated that hADSCs effectively facilitated photoreceptor regeneration and restoration of retinal function. By comparing the effects of isogenic bone marrow mononuclear stem cell (BM-MNC) transplantation on two animal models with different etiologies, namely RCS and P23H-1 rats, Di Pierdomenico et al. observed that intravitreous and subretinal homologous BM-MNC transplantation mitigated photoreceptor degeneration and exhibited anti-glioma.
properties; however, it did not enhance retinal function. Furthermore, isogenic BM-MNCs transplantation demonstrated superior efficacy compared to xenotransplantation of these cells. Given the potential risk of blood vessel obstruction from intravenous infusion cells becoming blocked in the lung, Liang et al.\(^\text{[46]}\) utilized a 10 μm filter to obtain small-cell UCMSCs (S-UCMSCs) and found that, compared to UCMSCs, intravenous infusion of S-UCMSCs is a safer option. In fact, it has been shown to delay retinal degeneration and protect visual function in RCS rats, making it a promising treatment method for RP. Liu et al.\(^\text{[50]}\) investigated various types of stem cells, including hADSCs, human amniotic fluid stem cells (hAFSCs), human bone marrow stem cells (hBMSCs), human dental pulp stem cells (hDPSCs), hiPSCs, and hiPSC-derived RPE cells, in their study on the protective effects and therapeutic potential of subretinal transplantation in rats with RP disease. They discovered that both adult and fetal stem cell injections resulted in improved visual function within 4wk, primarily attributed to the paracrine activity of multiple growth factors secreted by these stem cells.

**Clinic Trial** As for clinical trials, a variety of experiments have shown that stem cell therapy can alleviate clinical symptoms in patients with RP (Table 2).\(^\text{[51-57]}\). Kahraman and Oner\(^\text{[53]}\) conducted a clinical trial in which UCMSCs suspension was injected into transplanted fatty tissue between the choroid and sclera in 124 eyes of 82 patients during a six-month follow-up. Suprachoroidal administration of stem cells has beneficial effects on best corrected vision acuity (BCVA), visual field (VF) testing, and multifocal electroretinogram (mfERG) recording measurements. Zhao et al.\(^\text{[52]}\) conducted a comparative study to assess the safety and efficacy of modified subtenon capsule injection of triamcinolone alone (TA) and intravenous infusion of UCMSCs in the treatment of RP combined with macular edema (ME) (RP-ME). No serious adverse reactions were observed in all patients, indicating that both the modified sub-tenon capsule injection of TA and the intravenous infusion of UCMSCs are safe for RP-ME patients. In terms of effectiveness, TA injections demonstrated superior short-term outcomes in alleviating ME and improving visual function, while the effect of intravenous UCMSC on ME relief was slower but sustained over a longer period, leading to improved visual function\(^\text{[52]}\). The Mer tyrosine kinase (MERTK) gene encodes the Tyro3/Axl/Mer family of receptors, which are implicated in RP. Tagawa et al.\(^\text{[53]}\) successfully generated hiPSCs from RP patients and healthy individuals carrying homozygous or complex heterozygous mutations in MERTK. These hiPSCs were then differentiated into RPE cells. Although there were no significant morphological differences observed between diseased and normal RPE cells, cytokeratin staining indicated potential minor interference with cell polarity. Notably, the internalization of photoreceptor outer segments was significantly reduced in diseased hiPSC-RPE cells compared to normal hiPSC-RPE cells. This in vitro disease model holds promise for elucidating disease progression mechanisms and screening potential treatments. In 2009, a kind of original stem cell, a Wharton’s Jelly-derived mesenchymal stem cell, was extracted and cultured from the umbilical cord Wharton’s Jelly of the fetus after birth. Özmert and Arslan\(^\text{[54]}\) injected its suspension into the subtendon space of each patient’s eye. The research results showed that

### Table 2 Advantages and disadvantages of different stem cell therapies

<table>
<thead>
<tr>
<th>Types of stem cells</th>
<th>Methods</th>
<th>Advantages</th>
<th>Disadvantages</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>UCMSCs</td>
<td>Mesenchymal stem cell suspension was injected into transplanted fatty tissue between the choroid and sclera</td>
<td>Beneficial effects on BCVA, VF, and mfERG</td>
<td>Follow-up time was short; results do not apply to patients with early-stage disease</td>
<td>[51]</td>
</tr>
<tr>
<td>UCMSCs</td>
<td>Subtenon capsule injection of TA and intravenous infusion of UCMSCs</td>
<td>TA injections demonstrated superior short-term outcomes in improving visual function</td>
<td>The effect of intravenous UCMSC on ME relief was slower but sustained over a longer period</td>
<td>[52]</td>
</tr>
<tr>
<td>iPSCs</td>
<td>Differentiated iPSCs carrying homozygous or complex heterozygous mutations of MERTK into RPE</td>
<td>The internalization of photoreceptor outer segments was significantly reduced</td>
<td>Minor interference with cell polarity</td>
<td>[53]</td>
</tr>
<tr>
<td>WI-MSC</td>
<td>Injected its suspension into the subtendon space of each patient’s eye</td>
<td>An effective and safe option to slow or stop the progression of the disease</td>
<td>Intracellular mutant protein deposits could be detected by FAF; changes in deposits; FAF-field and visual field are not correlated in some cases</td>
<td>[54,57]</td>
</tr>
<tr>
<td>BMSCs</td>
<td>Conduct suprachoroidal implantation of MSCs in RP patients with relatively good vision</td>
<td>Effective in preventing apoptosis and promoting retinal tissue healing</td>
<td>Small sample size and short duration of follow-up; genetic factors and response to treatment were ignored</td>
<td>[55]</td>
</tr>
<tr>
<td>Allogeneic iPSCs</td>
<td>Performed allogeneic iPSC-derived retinal organoid transplantation in two patients with advanced RP</td>
<td>Visual function deterioration was observed to progress at a slower rate</td>
<td>Further investigation is required to assess its safety and efficacy in treating visual function</td>
<td>[56]</td>
</tr>
</tbody>
</table>

UCMSCs: Umbilical cord mesenchymal stem cells; BCVA: Best corrected visual acuity; VF: Visual field; mfERG: Multifocal electroretinogram; FAF: Fundus autofluorescence; TA: Triamcinolone alone; ME: Macular edema; iPSCs: Induced pluripotent stem cells; MERTK: Mer tyrosine kinase; RPE: Retinal pigment epithelium; BMSCs: Bone mesenchymal stem cells; WJ-MSC: Wharton’s Jelly-derived mesenchymal stem cells.
this transplant method can be considered an effective and safe option to slow or stop the progression of the disease, regardless of the gene mutation. By conducting suprachoroidal implantation of bone MSCs (BMSCs) in patients, Özkan et al. [55] observed that the implementation of spherical MSCs as a stem cell therapy in RP patients with relatively good vision led to improvements in BCVA, 10-2 and 30-2 VF examination, and mfERG records during the follow-up period. This suggests that using globular MSCs with enhanced effects may be more effective in preventing apoptosis and promoting retinal tissue healing in RP patients. Hirami et al. [56] conducted a study where they performed allogeneic hiPSC-derived retinal organoid transplantation in two patients with advanced RP. The results showed that the grafts remained viable for 2y, leading to increased retinal thickness at the graft site and no occurrence of serious adverse events in either patient. Moreover, during follow-up, visual function deterioration was observed to progress at a slower rate compared to untreated eyes. These findings highlight the potential therapeutic value of allogeneic hiPSC-derived retinal organoid transplantation; however, further investigation is required to assess its safety and efficacy in treating visual function [56]. The study conducted by Özmert and Arslan [57] demonstrated that the combination therapy of WJ-MSC and Magnovision can effectively decelerate the progression of RP in patients for a duration of up to 3y. These results suggest that stem cell therapy can significantly improve visual impairment in patients with RP and its safety has been confirmed by a variety of experiments. Therefore, stem cell therapy is widely used in clinical practice.

CONCLUSIONS
The treatment of RP primarily focuses on symptomatic management and retarding the degeneration of retinal cells. This includes drug therapy, nutritional support therapy, and the application of nerve growth factors. With advancements in medicine, gene therapy, tissue therapy, stem cell therapy, and other approaches have been proposed. Unfortunately, gene therapy can only target a single gene and is effective for a limited number of patients. Tissue therapy may lead to immune rejection and potential tumorigenic risks. Due to their self-renewal ability and multiple differentiation potentials, stem cells offer hope for curing RP. Extensive innovative animal and clinical studies provide a strong foundation for the feasibility and safety of stem cell therapy in RP patients, thus making it one of the most effective treatment options available [58]. Stem cells can fully regenerate RPE cells to replace those that have lost function. Furthermore, stem cell transplantation offers several advantages, such as protecting retinal blood vessels, promoting the repair of damaged nerve cells, nourishing nerves, inhibiting inflammatory responses, etc. However, it should be noted that stem cell therapy also has certain limitations, including variability in animal models and human mutations, immature gene transfer techniques, surgical complications, and local or systemic immunosuppression complications. Additionally, its long-term efficacy remains uncertain. At present, placental MSCs show broad prospects in the treatment of RP, but they are still poorly understood and need further research [59].

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