· Commentary ·

TRPC6: an underlying target for human glaucoma

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Abstract

· Glaucoma is one of the leading causes of visual impairment and blindness worldwide. Of known risk factors for glaucoma, an increased in intraocular pressure is most highly correlated with glaucomatous damage. Irrespective of the cause, apoptosis of the retinal ganglion cells is the eventual outcome. It is widely accepted that glaucoma is a neurodegenerative disease that is strongly correlated with central nervous system disorders, such as Alzheimer's disease. These two disorders also share some similarities in pathogenic mechanisms. Recent studies suggest that the transient receptor potential canonical 6 channel could work together with brain-derived neurotrophic factor to promote neuron survival in brain and retina. In this study, we propose that transient receptor potential canonical 6 may contribute to the pathogenesis of human glaucoma and become a potential therapeutic target.

• KEYWORDS: glaucoma; transient receptor potential canonical 6 channel; neuroprotection

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INTRODUCTION

R etinal ganglion cells (RGCs) death is the final step in glaucoma and other ocular neurodegenerative diseases ^[1]. RGCs have been considered as an attractive reciprocal model to study neurodegenerative disorders in the central nervous system (CNS)^[2-4]. At present, the exact mechanisms of RGCs death remain unknown, and there is no effective treatment to combat cell death. The initiation and progression of glaucomatous optic neuropathy and

preferential killing of RGCs attributed to elevated intraocular pressure (IOP)^[5]. Insufficient blood perfusion caused by increased IOP or ischemia of the optic nerve head may also cause progressive RGCs death with resultant degeneration of the optic nerve and, ultimately, vision loss ^[6]. Currently, primary treatment of glaucoma is aimed at lowering the IOP^[7]. However, some individuals with significantly lowered IOP following treatment show no improvement in glaucomatous optic neuropathy and even progression of the disease [8]. Other factors may, therefore, be implicated in RGCs death, and it is likely that neuroprotection, in addition to lowering of IOP, needs to be considered in glaucoma treatment. Previous studies have demonstrated the potent role of brain-derived neurotrophic factor (BDNF) in promoting neurons survival, including RGCs ^[9-15]. Recent studies have also revealed that transient receptor potential canonical 6 (TRPC6) participates in BDNF-mediated neuron survival in the CNS and calcium (Ca^{2+}) entry may be involved in early onset Alzheimer's disease (AD) [16,17]. Thus, TRPC6 may contribute to neurodegenerative changes. The potential link between glaucoma and AD has already been identified, with a previous study reporting a high rate of glaucoma among patients with AD ^[18]. We propose that TRPC6 may play a critical role in signaling cascades involving RGCs survival in glaucoma. BDNF-mediated RGCs survival and TRPC6mediated Ca²⁺ regulation may be key aspects of the process. Elucidating the potential role of TRPC6 in glaucoma may vield possible therapeutic targets for the treatment of glaucomatous optic neuropathy.

TRANSIENT RECEPTOR POTENTIAL AND TRANSIENT RECEPTOR POTENTIAL CANONICAL Transient receptor potential (TRP) channels are cationpermeable channels first identified in phototransduction mutation studies of *Drosophila*^[18,19]. More than 30 members of the TRP family have been cloned in both vertebrates and invertebrates. TRPs are broadly expressed in a variety of organisms and mediate multiple physiological functions^[20-22] such as neurotransmitter release, neurite outgrowth, cell cycle regulation, cell apoptosis and survival via the regulation of Ca²⁺ changes ^[21,23]. TRPs are also expressed in the transduction of various responses such as mechanosensation ^[24]. Among the mammalian TRP superfamily, TRP canonical (TRPC) channels show the most structural similarity to the *Drosophila* TRPs ^[25]. The TRPC family (TRPC1-7) can be divided by homology and function into four subfamilies: TRPC1, TRPC2, TRPC4/5, and

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TRPC3/6/7 ^[21]. The roles of TRPC6 are explained based on the following aspects.

EXPRESSION AND FUNCTION OF TRPC6

TRPC6 is widely expressed in mammalian brain and retina. In rat brain, TRPC6 was reported in dentate gyrus in the hippocampus ^[25], cerebellar granule neurons (CGNs) in the cerebellum ^[16], and substantia nigra in the midbrain ^[26]. In retina, TRPC6 was expressed in RGCs ^[27-29], rods ^[30], and many other cell types^[31,32].

TRPC6 is a key player in neuron pathophysiological functions ^[16, 33-35]. It was essential in BDNF-mediated neuron growth cone turning and intracellular Ca²⁺ elevation ^[33]. Down-regulation of TRPC6 led to apoptosis and blocked the BDNF-protective effect in CGNs, and overexpression of TRPC6 could protect CGNs against serum deprivation-induced cell death ^[16]. TRPC6 promoted neuron dendritic growth via the CaMKIV-CREB pathway ^[34], which suggested that TRPC6 was important during brain development ^[35].

In retina, preliminary work has been conducted in TRP channels research. Wang *et al*^{(36,37]} showed that constitutively active TRPCs resulted in retinal degeneration because of Ca²⁺ overload. TRPC channel blockers can suppress light-evoked currents in rat photosensitive RGCs ^[28]. TRPC may also mediate basal Ca²⁺ entry in retinal pigment epithelial (RPE) cells ^[32] and Müller cells ^[31]. Activating TRPC6 channel prior to ischemia has early neuroprotective effects on RGCs *in viva* and the protection of TRPC6 was also BDNF-mediated ^[29].

As a potent neuroprotective agent, BDNF appears to be a good candidate for therapeutic treatment for some CNS disorders such as AD ^[38,39]. BDNF-dependent pathways of neuron survival in the CNS may share similarities with that of RGCs in retina. In fact, BDNF had long been known to influence RGCs survival, both during retinal development and following lesioning ^[9-13]. TRPC has been clarified as the key downstream target for the neuronal protective effect of BDNF ^[16]. TRPC6 is required for the neuroprotective effect of BDNF.

POTENTIAL DISEASE-RELATED MECHANISMS

The potential link between AD and glaucoma indicates that TRPC6 might be involved the pathogenesis of glaucoma. Ca^{2+} is a ubiquitous and versatile intracellular messenger, which regulates many cellular activities such as neuronal excitability, synaptic plasticity, and neurotoxicity. Dysfunction of Ca^{2+} homeostasis can result in many CNS-related cognitive and neuropathological problems.

AD is a chronic neurodegenerative disease. Although the molecular basis has not been fully established ^[40,41], TRPC6-induced dysfunction of Ca²⁺ homeostasis may be the key step. It is generally accepted that abeta deposits, which renders neurons vulnerable to excitotoxicity and apoptosis by disruption of intracellular Ca²⁺ homeostasis and production of neurotoxic factors, are central to its neuropathology ^[42]. Previous report showed that the

overexpression of presenilin-2 and AD-linked presenilin-2 variants influenced TRPC6-enhanced Ca^{2+} entry into HEK293 cells. Thus, TRPC6 in AD may be a promising focus for research^[43].

A high prevalence of glaucoma had been found in AD patients ^[44-46], with many AD patients exhibiting typical glaucomatous optic neuropathy, such as enlarged optic disc cupping, damaged retinal nerve fibers, and a defective visual field. To some extent, glaucoma had been proposed to be a form of ocular AD ^[1]. Therefore, it is reasonable to believe that the underlying pathways of the two diseases may be similar. Future studies may reveal a similar mechanism of TRPC6 activity in glaucoma and AD.

TRPC6, PRESSURE AND GLAUCOMA

TRPC6 is also associated with pressure-related changes in many diseases. High TRPC6 levels have been found in numerous tissues including the lung, stomach, colon, esophagus, and myometrium ^[47]. TRPC6 can be directly activated by mechanical stimulation ^[48,49]. The intravascular pressure-induced depolarization and constriction of small arteries and arterioles was regulated by TRPC6^[50]. Moreover, TRPC6 expression was up-regulated in pulmonary arteries hypertension of rats ^[51]. Podocin, together with TRPC6 and possibly other TRPCs, forms complexes with other transmembrane proteins may act to sense glomerular pressure ^[52]. These characteristics of activation make TRPC6 serving as an environmental pressure sensor, translating extracellular cues into intracellular signals.

Glaucoma is a typical optic neuropathy with IOP as the main risk factor. The pressure inducing RGCs death remains unclear. Designed to sense pressure, a series of receptors are located throughout the brain and retina pathway. These molecules might allow cells in the retina and optic nerve to respond directly to ocular pressure. Pressure injury may overload the cells with Ca²⁺, which cause a direct degenerative cascade. The molecules sensing pressure might be the factor translating pressure into neuronal damage. TRPV1, another member of the TRP family, one of the sensor of pressure, has recently been discovered expressed in RGCs and induced apoptosis through the influx of extracellular Ca²⁺ ^[53]. The possibility that TRPC6, also a pressure-sensitive channel, may exert similar functions or work together is worth investigation.

HYPOTHESIS

Taking the features of TRPC6 already mentioned above into consideration, we hypothesize that TRPC6 may contribute to the pathogenesis of human glaucoma and become a potential therapeutic target. Thus manipulation of TRPC6 activity may provide a novel and promising tool for the treatment of the disease.

In conclusion, glaucoma is a major cause of optic neuropathy and adversely affects the vision and life quality of many people in a large number of developing countries^[54,55]. At present, there is no effective treatment strategy to protect against optic nerve damage induced by intraocular hypertension and other risk factors. There has been little success in the development of neuroprotective agents for retinal damage. Firstly, there is the issue of timing: a drug would have to be administered within a reasonable time of the retinal insult occurring ^[56]. Secondly, all drugs that have been developed have been characterized by incomplete protection and presented a risk of nonspecific effects, such as glutamate excitotoxicity ^[57] or other side effects. Thus far, most current studies have focused on reducing IOP. However, targeting IOP lowering will not completely impede the progression of glaucoma in all patients. Neuroprotection has, therefore, become increasingly important as a therapeutic target, with innovative studies underway [58]. Experimental data previously revealed neuroprotective effects of various agents and strategies, such as neurotrophin delivery^[59] and blocking of excess glutamate stimulation ^[60]. Other strategies attempted include stabilization of Ca²⁺ homeostasis ^[61], prevention of apoptosis, modulation of immunologic status *via* vaccination ^[62], and induction of endogenous neuroprotective mechanisms. These studies raise hopes for discovering beneficial effects in future clinical trials.

The potential role of TRPC6 as a neuroprotective target is illustrated as follows: 1) The possibility which TRPC6 is involved in neurodegenerative diseases is a reasonable conjecture. Identifying the physiological signals that regulate TRPC6 activity in glaucoma appears to be a clear priority. To date, little work has been done in this area. TRPC6 would become an important and interesting target in glaucoma research; 2)TRPC6 appears to be important in the pathogenetic pathway that leads to apoptosis of RGCs ^[29]. In different retinal cell types, we could discover whether TRPC6 plays an active role in Ca2+ entry pathways. If TRPC6 channel were found to control a variety of biological functions, new and promising drug development could emerge; 3)Targeting TRPC6 may be helpful in protecting RGCs against elevated IOP and other insults. The damage to RGCs occurs at an early stage of glaucoma, even before visual field defects are detected. In general, pressure-induced dysfunction of RGCs precedes cell death; therefore, neuroprotective therapies could be more effective at this stage. Our hypothesis suggests a possible method to detect glaucoma at an early stage and monitor the development of the disease; 4) TRPC6 may enhance our understanding of the mechanisms of RGCs neurodegeneration and provide new insight in optic neuropathy, as well as other neurodegenerative diseases, such as AD. Overall, exciting advances at the laboratory level will continue to drive research on the role of TRPC6 in glaucoma. Future investigations of human glaucoma and AD involving TRPC6 should prove highly rewarding in the years to come.

2003;8:s1140-156

2 Weishaupt JH, Bähr M. Degeneration of axotomized retinal ganglion cells as a model for neuronal apoptosis in the central nervous system-molecular death and survival pathways. *Restor Neurol Neurosci* 2001;19 (1-2):19-27

3 Guerin MB, McKernan DP, O'Brien CJ, Cotter TG. Retinal ganglion cells: dying to survive. *Int J Dev Biol*2006;50(8):665-674

4 Ning A, Cui J, To E, Ashe KH, Matsubara J. Amyloid-beta deposits lead to retinal degeneration in a mouse model of Alzheimer disease. *Invest Ophthalmol Vis Sci*2008;49(11):5136-5143

5 Nickells RW. From ocular hypertension to ganglion cell death: a theoretical sequence of events leading to glaucoma. *Can J Ophthalmol.* 2007;42(2):278-287

6 Dahlmann-Noor AH, Vijay S, Limb GA, Khaw PT. Strategies for optic nerve rescue and regeneration in glaucoma and other optic neuropathies. *Drug Discov Today* 2010;15(7-8):287-299

7 Weinreb RN. Glaucoma neuroprotection: why is it? Why is it needed? *Can J Ophthalmol* 2007;42(3):396-398

8 Mozaffarieh M, Flammer J. Is there more to glaucoma treatment than lowering IOP? *Surv Ophthalmol* 2007;52(Suppl 2):S174-179

9 Johnson JE, Barde YA, Schwab M, Thoenen H. Brain-derived neurotrophic factor (BDNF) supports the survival of cultured rat retinal ganglion cells. *JNeurosci*1986;6(10):3031–3038

10 Arango-González B, Cellerino A, Kohler K. Exogenous brain-derived neurotrophic factor (BDNF) reverts phenotypic changes in the retinas of transgenic mice lacking the BDNF gene. *Invest Ophthalmol Vis Sci* 2009; 50(3):1416-1422

11 Cooper NG, Laabich A, Fan W, Wang X. The relationship between neurotrophic factors and CaMKII in the death and survival of retinal ganglion cells. *Prog Brain Res*2008;173:521-540

12 Parrilla-Reverter G, Agudo M, Sobrado-Calvo P, Salinas-Navarro M, Villegas-Pérez MP, Vidal-Sanz M. Effects of different neurotrophic factors on the survival of retinal ganglion cells after a complete intraorbital nerve crush injury: a quantitative *in vivo*study. *Exp Eye Res*2009;89(1):32-41

13 Chen H, Weber AJ. BDNF enhances retinal ganglion cell survival in cats with optic nerve damage. *Invest Ophthalmol Vis Sci* 2001;42 (5): 966–974

14 Spalding KL, Rush RA, Harvey AR. Target-derived and locally derived neurotrophins support retinal ganglion cell survival in the neonatal rat retina. *J.Neurophiol* 2004;60(3):319–327

15 Bonnet D, Garcia M, Vecino E, Lorentz JG, Sahel J, Hicks D. Brain-derived neurotrophic factor signalling in adult pig retinal ganglion cell neurite regeneration *in vitra Brain Res*2004;1007(1-2):142-151

16 Jia YC, Zhou J, Tai YL, Wang YZ. TRPC channels promote cerebellar granule neuron survival. *Nat Neurosci* 2007;10(5):559–567

17 Lessard CB, Lussier MP, Cayouette S, Bourque G, Boulay G. The overexpression of presenilin2 and Alzheimer's-disease-linked presenilin2 variants influences TRPC6-enhanced Ca2+ entry into HEK293 cells. *Ccll Signal* 2005;17(4):437-445

18 Minke B. Drosophila mutant with a transducer defect. *Biophys Struct Mech*1977;3(1):59-64

19 Montell C, Jones K, Hafen E, Rubin G. Rescue of the Drosophila phototransduction mutation trp by germline transformation. *Science* 1985; 230(4729):1040-1043

20 Montell C, Birnbaumer L, Flockerzi V. The TRP channels, a remarkably functional family. *Cell*2002;108(5):595–598

21 Nilius B, Owsianik G, Voets T, Peters JA. Transient receptor potential cation channels in disease. *Physiol Rev*2007;87(1):165–217

22 Talavera K, Nilius B, Voets T. Neuronal TRP channels: thermometers, pathfinders and life-savers. *Trends Neurosci*2008;31(6):287-295

23 Voets T, Talavera K, Owsianik G, Nilius B. Sensing with TRP channels.

1 McKinnon SJ. Glaucoma: ocular Alzheimer's disease? Front Biosci

REFERENCES

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Nature Chem Bio/2005;1(2):85-92

24 Clapham DE. TRP channels as cellular sensors. *Nature* 2003;426 (6966):517-524

25 Putney JW. Physiological mechanisms of TRPC activation. *Pflugers* Arch2005;451(1):29-34

26 Giampá C, DeMarch Z, Patassini S, Bernardi G, Fusco FR. Immunohistochemical localization of TRPC6 in the rat substantia nigra. *Neurosci Lett* 2007;424(3):170–174

27 Warren EJ, Allen CN, Brown RL, Robinson DW. The light-activated signaling pathway in SCN-projecting rat retinal ganglion cells. *Eur J Neurosci*2006;23(9):2477-2487

28 Sekaran S, Lall GS, Ralphs KL, Wolstenholme AJ, Lucas RJ, Foster RG, Hankins MW. 2-Aminoethoxydiphenylborane is an acute inhibitor of directly photosensitive retinal ganglion cell activity *in vitro* and *in viva J* Neurosci2007;27(15):3981-3986

29 Wang X, Teng L, Li A, Ge J, Laties AM, Zhang X. TRPC6 channel protects retinal ganglion cells in a rat model of retinal ischemia/ reperfusion-induced cell death. *Invest Ophthalmol Vis Sci* 2010;51(11): 5751-5758

30 Krizaj D. Compartmentalization of calcium entry pathways in mouse rods. *Eur J Neurosci* 2005;22(12):3292-3296

31 Da Silva N, Herron CE, Stevens K, Jollimore CA, Barnes S, Kelly ME. Metabotropic receptor-activated calcium increases and store-operated calcium influx in mouse Müller cells. *Invest Ophthalmol Vis Sci* 2008;49 (7):3065–3073

32 Wimmers S, Strauss O. Basal calcium entry in retinal pigment epithelial cells is mediated by TRPC channels. *Invest Ophthalmol Vis Sci* 2007;48(12):5767–5772

33 Li Y, Jia YC, Cui K, Li N, Zheng ZY, Wang YZ, Yuan XB. Essential role of TRPC channels in the guidance of nerve growth cones by brain-derived neurotrophic factor. *Nature*2005;434(7035):894-898

34 Tai YL, Feng SJ, Ge RL, Du WL, Zhang XX, He ZH, Wang Y. TRPC6 channels promote dendritic growth via the CaMKIV-CREB pathway. *J Cell Sci*2008;121(Pt 14):2301-2307

35 Tai Y, Feng S, Du W, Wang Y. Functional roles of TRPC channels in the developing brain. *Pflugers* 4rch2009;458(2):283-289

36 Wang T, Xu H, Oberwinkler J, Gu Y, Hardie RC, Montell C. Light activation, adaptation, and cell survival functions of the Na⁺/Ca²⁺ exchanger CalX. *Neuron*2005;45(3):367–378

37 Wang T, Montell C. Phototransduction and retinal degeneration in Drosophila. *Pflugers Arch*2007;454(5):821-847

38 Pezet S, Malcangio M. Brain-derived neurotrophic factor as a drug target for CNS disorders. *Expert Opin Ther Targets*2004;8(5):391-399

39 Nagahara AH, Merrill DA, Coppola G, Tsukada S, Schroeder BE, Shaked GM, Wang L, Blesch A, Kim A, Conner JM, Rockenstein E, Chao MV, Koo EH, Geschwind D, Masliah E, Chiba AA, Tuszynski MH. Neuroprotective effects of brain-derived neurotrophic factor in rodent and primate models of Alzheimer's disease. *Nat Mcd* 2009;15(3):331-337

40 Yin H, Chen L, Chen X, Liu X. Soluble amyloid beta oligomers may contribute to apoptosis of retinal ganglion cells in glaucoma. *Med Hypotheses*2008;71(1):77-80

41 Vasto S, Candore G, Listì F, Balistreri CR, Colonna-Romano G, Malavolta M, Lio D, Nuzzo D, Mocchegiani E, Di Bona D, Caruso C. Inflammation, genes and zinc in Alzheimer's disease. *Brain Res Rev* 2008; 58(1):96-105

42 Huang HC, Jiang ZF. Accumulated amyloid-beta peptide and hyperphosphorylated tau protein: relationship and links in Alzheimer's disease. *J Alzheimers Dis*2009;16(1):15-27

43 Yamamoto S, Wajima T, Hara Y, Nishida M, Mori Y. Transient receptor potential channels in Alzheimer's disease. *Biochim Biophys Acta* 2007; 1772(8):958-967 44 Tamura H, Kawakami H, Kanamoto T, Kato T, Yokoyama T, Sasaki K, Izumi Y, Matsumoto M, Mishima HK. High frequency of open-angle glaucoma in Japanese patients with Alzheimer's disease. *J Neurol Sci* 2006;246(1-2):79-83

45 Bayer AU, Keller ON, Ferrari F, Maag KP. Association of glaucoma with neurodegenerative diseases with apoptotic cell death: Alzheimer's disease and Parkinson's disease. *Am J Ophthalmol*2002;133(1):135–137

46 Bayer AU, Ferrari F, Erb C. High occurrence rate of glaucoma among patients with Alzheimer's disease. *Eur Neurol*2002;47(3):165-168

47 Beech DJ, Muraki K, Flemming R. Non-selective cationic channels of smooth muscle and the mammalian homologues of Drosophila TRP. / *Physiol*2004;559(Pt 3):685-706

48 Sharif-Naeini R, Dedman A, Folgering JH, Duprat F, Patel A, Nilius B, Honoré E. TRP channels and mechanosensory transduction: insights into the arterial myogenic response. *Pflugers* 4rch2008;456(3):529-540

49 Spassova MA, Hewavitharana T, Xu W, Soboloff J, Gill DL. A common mechanism underlies stretch activation and receptor activation of TRPC6 channels. *Proc Natl Acad Sci USA*2006;103(44):16586-16591

50 Welsh DG, Morielli AD, NelsonMT, Brayden JE. Transient receptor potential channels regulate myogenic tone of resistance arteries. *Circ Rcs* 2002;90(3):248–250

51 Lin MJ, Leung GP, Zhang WM, Yang XR, Yip KP, Tse CM, Sham JS. Chronic hypoxia-induced upregulation of store-operated and receptoroperated Ca²⁺ channels in pulmonary arterial smooth muscle cells: a novel mechanismofhypoxic pulmonary hypertension. *Circ Res* 200495(5):496-505

52 Huber TB, Schermer B, Müller RU, Höhne M, Bartram M, Calixto A, Hagmann H, Reinhardt C, Koos F, Kunzelmann K, Shirokova E, Krautwurst D, Harteneck C, Simons M, Pavenstädt H, Kerjaschki D, Thiele C, Walz G, Chalfie M, Benzing T. Podocin and MEC-2 bind cholesterol to regulate the activity of associated ion channels. *Proc Natl Acad Sci US* 12006;103(46):17079-17086

53 Sappington RM, Sidorova T, Long DJ, Calkins DJ. TRPV1: Contribution to retinal ganglion cell apoptosis and increased intracellular Ca²⁺ with exposure to hydrostatic pressure. *Invest Ophthalmol Vis Sci* 2009;50(2):717–728

54 Cedrone C, Mancino R, Cerulli A, Cesareo M, Nucci C. Epidemiology of primary glaucoma: prevalence, incidence, and blinding effects. *Prog Brain Res*2008;173:3-14

55 Thomas R, Sekhar GC, Parikh R. Primary angle closure glaucoma: a developing world perspective. *Clin Experiment Ophthalmo*/2007;35(4): 374-378

56 Levin LA, Peeples P. History of neuroprotection and rationale as a therapy for glaucoma. *Am J Manag Carc* 2008;14(1 Suppl):S11-14

57 Seki M, Lipton SA. Targeting excitotoxic/free radical signaling pathways for therapeutic intervention in glaucoma. *Prog Brain Res* 2008; 173:495-510

58 Bron A. Neuroprotection: is it close to us? JFr Ophtalmol 2009;32(3): 217–220

59 Fujino H, Kitaoka Y, Hayashi Y, Munemasa Y, Takeda H, Kumai T, Kobayashi S, Ueno S. Axonal protection by brain-derived neurotrophic factor associated with CREB phosphorylation in tumor necrosis factor-alpha-induced optic nerve degeneration. *Acta Neuropathol* 2009; 117(1):75-84

60 Fang JH, Wang XH, Xu ZR, Jiang FG. Neuroprotective effects of bis(7) -tacrine against glutamate-induced retinal ganglion cells damage. *BMC Neurosci*2010;11:31

61 Wang RF, Gagliuso DJ, Podos SM. Effect of flunarizine, a calcium channel blocker, on intraocular pressure and aqueous humor dynamics in monkeys. *J Glaucoma*2008;17(1):73-78

62 Schwartz M. Modulating the immune system: a vaccine for glaucoma? Can J Ophthalmol 2007;42(3):439-441