• Basic Research •

Two mutations in the transforming growth factor betainduced gene associated with familial Lattice corneal dystrophy

Wen-Ping Cao¹, Hai-Gang Yuan², Ping Liu¹, Xue Li¹, Qi Hu¹

¹Eye Hospital, the First Affiliated Hospital, Harbin Medical University, Harbin 150001, Heilongjiang Province, China ²First Affiliated Hospital, Heilongjiang University of Chinese Medicine, Harbin 150001, Heilongjiang Province, China **Co-first authors**: Wen-Ping Cao and Hai-Gang Yuan **Correspondence to**: Qi Hu. Eye hospital, the First Affiliated Hospital, Harbin Medical University, Harbin 150001, Heilongjiang Province, China. huqi51115@sina.com

Received: 2016-07-27 Accepted: 2016-11-05

Abstract

• AIM: To report a phenotypic variant pedigree of lattice corneal dystrophy (LCD) associated with two mutations, R124C and A546D, in the transforming growth factor beta-induced gene (TGFBI).

• METHODS: A detailed ocular examination was taken for all participants of a LCD family. Peripheral blood leukocytes from each participant were extracted to obtain the DNA. Polymerase chain reaction (PCR) of all seventeen exons of TGFBI gene was performed. The products were sequenced and analyzed. Histological examination was carried out after a penetrating keratoplasty from the right eye of proband.

• RESULTS: Genetic analysis showed that the proband and all 6 affected individuals harbored both a heterozygous CGC to TGC mutation at codon 124 and a heterozygous GCC to GAC mutation at codon 546 of TGFBI. None of the 100 control subjects and unaffected family members was positive for these two mutations. Ocular examination displayed multiple refractile lattice-like opacities in anterior stroma of the central cornea and small granular deposits in the peripheral cornea. The deposits were stained positively with Congo red indicating be amyloid in nature and situated mainly in the anterior and middle stroma.

• CONCLUSION: We observed a novel LCD family which carried two pathogenic mutations (R124C and A546D) in the TGFBI gene. The phenotypic features were apparently different from those associated with corresponding single mutations. The result reveals that although the definite mutation is the most important genetic cause of the disease, some different modifier alleles may influence the phenotype. • **KEYWORDS:** corneal dystrophy; mutation; phenotype; transforming growth factor beta-induced gene **DOI:10.18240/ijo.2017.03.03**

Cao WP, Yuan HG, Liu P, Li X, Hu Q. Two mutations in the transforming growth factor beta-induced gene associated with familial Lattice corneal dystrophy. *Int J Ophthalmol* 2017;10(3):343-347

INTRODUCTION

T GFBI (OMIM 601692, formerly called BIGH3) gene was first identified by Skonier *et al*^[1] as a transforming growth factor beta-induced gene in a human lung adenocarcinoma cell line. In 1997, Munier *et al*^[2] identified TGFBI on chromosome 5q31 and discovered 4 different mutations that associated with 4 inherited corneal dystrophies: R555W resulting in the granular dystrophy (GCD), R124H resulting in the Avellino dystrophy (ACD), R124C resulting in the lattice dystrophy type I (LCD-I), R555Q resulting in the Reis-Bücklers dystrophy (RBCD).

Subsequently, several additional mutations of TGFBI throughout the world were found to be responsible for diverse corneal dystrophies and a phenotype-genotype correlation had been established^[3-4].

LCD is one of common stromal dystrophy which manifests typically as linear, radially oriented, branching opacities in the anterior stroma^[5]. The opacities have been found to correspond with accumulations of amyloid. At least four different types of LCDs are recognized based on clinical features and the histologically natures. But it is difficult to distinguish these subtypes from each other in the absence of genetic analysis. Mutations in the TGFBI gene have been found in patients with LCD type I and IIIA. Mutations in the GSN gene have been found in cases of LCD type II^[6-7]. The autosomal recessive form of the disease, LCD type III, has been mapped to chromosome 1p31^[8]. To date, several distinct single mutations of TGFBI which including R124C, A546D, P501T, L527R, A620P, L518P and H626R have been associated with LCD^[3,9-13]. It seems that these single nucleotide changes are important disease-producing mutations.

Several researchs have observed patients who harbored two different mutations in TGFBI with variant phenotype^[14-15].

	Table 1	Clinical	findings	and	genotype	in	family	members
--	---------	----------	----------	-----	----------	----	--------	---------

Individual case	Gender	Age (a)	Status	Mutations in TGFBI	Age of	Symptoms of	Visual acuity	
					onset (a)	onset	OD	OS
I 2	F	90	Affected	R124C	19-20	FBS; Ph	10 cm/CF	10 cm/CF
				A546D				
Ⅲ 2	М	56	Unaffected	None	-	-	0.8	1.0
Ⅲ 3	F	63	Affected	R124C	15	FBS; Ph; VA \downarrow	20 cm/CF	20 cm/CF
				A546D				
∐ 4	F	61	Affected	R124C	17	FBS; Ph; VA \downarrow	0.01	20 cm/CF
				A546D				
Ⅲ 2	М	28	Unaffected	None	-	-	1.0	1.0
	М	41	Affected	R124C	18	FBS; Ph	0.4	0.4
Ⅲ 3				A546D				
Ⅲ 4	М	39	Unaffected	None	-	-	1.0	1.0
Ⅲ 5	М	33	Affected	R124C	21	FBS; Ph	0.6	0.6
				A546D				
Ⅲ 6	F	39	Unaffected	None	-	-	1.0	1.0
	F	16	Affected	R124C	16	FBS	0.8	0.8
11 3				A546D				

OD: Right eye; OS: Left eye; FBS: Foreign body sensation; VA↓: Visual acuity decreased; Ph: Photophobia. -: Information is not provided.

Nevertheless, the reason of the phenotype in patients with such a double mutations differs from that with single mutations is still unclear. In this study, we report the first cases of a LCD family in Chinese associated with both R124C and A546D mutations of TGFBI gene.

SUBJECTS AND METHODS

Subjects The research strictly followed the Declaration of Helsinki and was adhered to the Review Board of Heilongjiang. A four-generation LCD pedigree was collected in the north of China in March 2014 (Figure 1). All 17 participants (6 affected and 11 unaffected) provided informed written consents. The ages of family members ranged from 5 to 90y. In addition, 100 unrelated, unaffected, healthy volunteers from China were collected as normal controls. The ages of normal controls ranged from 28 to 58y.

Clinical Evaluations Extensive ophthalmologic examinations were performed to determine the status of the corneas of each individual (affected or unaffected). The corneal phenotypes of affected members were documented by slit-lamp photography. The clinical history that including the age of onset, innitial presenting signs, clinical symptoms, and the treatment procedures were obtained in detail.

Molecular Analysis Peripheral blood was obtained in a total of 17 family members and one hundred volunteers. DNA was extracted with a QIAamp DNA blood Mini Kit (Qiagen, Hilden, Germany). Polymerase chain reaction (PCR) amplification of all 17 exons of TGFBI gene were performed using the appropriate primers and conditions previously reported by Munier *et al*^[2] or Afshari *et al*^[16]. Each PCR product was sequenced with ABI BigDye Terminator Cycle Sequencing



Figure 1 The pedigree of LCD family The proband was indicated by the arrow. Affected members in the family were filled with black and individuals who underwent research were indicated by the asterisks.

kit v3.1, (ABI Applied Biosystems, Foster City, CA, USA) for both strands. Then the results of sequencing were compared with the cDNA of TGFBI in GenBank (NM-000358) in order to detect any nucleotide changes. All Chinese controls were amplified and scanned the exon 4 and exon 12 of TGFBI gene. **Histologic Evaluations** One corneal button from the right eye of the proband was obtained after penetrating keratoplasty. Corneal section was stained by hematoxylin-eosin (HE), Congo red, periodic acid Schiff (PAS) and the Masson's trichrome for histologic examination.

RESULTS

Clinical Manifestations Clinical findings of the LCD pedigree were summarized (Table 1). Age of onset was 15 to 21y in the six affected family members. The initial symptoms and main complains included visual acuity decreased, foreign body



Figure 2 Slit-lamp photography of patients with LCD that associate with both R124C and A546D A: Photography of the proband showed gray and white confluent opacities involved almost whole cornea of her right eye; B: Clinical penotype of IV 3 (granddaughter of the proband) revealed several slight opacities in superficial stroma of the right corneal; C, D: Multiple refractile lattice-like opacities were finded in anterior stroma of the central cornea and small granular deposits could be seen in the peripheral cornea of III 3 and III 5 in their right eyes.

sensation and photophobia. Corneal erosion appeared in III 3 and III 5 in their third decade of life. The progress of corneal opacities in all affected members was commonly bilateral symmetry.

The proband of the family was a sixty-three years old Chinese woman who came to hospital for corneal transplantation. She complained a long-term bilateral decrease of visual acuity. Slitlamp examination revealed gray and white confluent opacities involved almost entire cornea (Figure 2A). Severe corneal edema presented and the typical lattice opacities could not be observed. Her visual acuity decreased to 20 cm/CF in both eyes. Ultimately, penetrating keratoplasty was performed on her both eyes respectively.

III 3 and III 5 had their initial symptoms at 18 and 21y and both of them complained with foreign body sensation and photophobia. Slit-lamp examination observed that multiple refractile lattice-like opacities in anterior stroma of the central cornea and small granular deposits could be seen in the peripheral cornea (Figure 2C, 2D). The visual acuity of both eyes decreased to 0.4 in III 3 and 0.6 in III 5 respectively.

IV 3 was the granddaughter of proband (Figure 2B). No severe symptom or visual defect was complained until participating this research. Clinical penotype showed that several slight opacities in superficial stroma of her right cornea.

Molecular Genetic Analysis Direct sequencing was taken for all exons of TGFBI gene from the proband, and we identified two heterozygous mutations (Figure 3A, 3C). The exon 4 exhibits a C > T heterozygous substitution at nucleotide position 417 and causes a arginine to cystine acid variation at protein level (R124C). The exon 12 exhibits a C > A heterozygous substitution at the 1637 position and results in a alanine to aspartic acid variation at protein level (A546D). Additionaly, 5 affected (I2, II4, III3, III5 and IV 3) individuals who carrying the same two variations in heterozygous state were identified. None of 11 unaffected (II2, II6, II7, II8, III2, III4, III6, III9, III10, IV 4 and IV 6) participants and the one hundred normal controls was positive for these two mutations (Figure 3B, 3D). Histologic Examination The proband underwent a penetrating keratoplasty for her right eye and the corneal button was excised for further histologic examination. It characterized by presence of variably sized, irregularly shaped deposits within the cornea. The deposits were stained positively with Congo red indicating be amyloid in nature and situated mainly in the anterior and middle stroma (Figure 4).

DISCUSSION

The authors presented the first report of a LCD pedigree which associated with two missense mutations in TGFBI gene, R124C in exon 4 and A546D in exon 12. These two heterozygous mucleotide changes not only cosegregated with keratopathy in the family, but also were predicted to change two amino acids of TGFBI-induced protein. It seems that both of these changes were important pathogenic mutations. As reported^[17], the typical LCD caused by R124C was recognized by the characteristic net of linear opacities in the anterior stroma. The patients often had signs and symptoms at the first decade of their lives and the recurrent epithelial erosion was common. However, the atypical LCD caused by A546D presented polymorphic, chipped ice-appearing corneal opacities or combined with filamentous opacities in the peripheral cornea^[18-19]. The amyloid deposits were larger and situated predominantly in the mid and posterior corneal stroma. They often had a later onset and rarely complain of corneal erosions. In the current studied family, the age at onset for affected individuals was between 15 and 21y, with a mean of 17.75y. In contrast to the typical LCD, mild corneal erosion only appeared in III 3 and III 5. Multiple refractile lattice-like opacities were presented in their anterior stroma of central cornea and small granular deposits were observed in peripheral cornea. Histologic result showed that the amyloid deposits situated mainly in the anterior and middle stroma compared with the atypical LCD. These clinical features of studied patients were apparently different from those associated with corresponding single mutations. At this point of view, the phenotype of our pedigree was seemed to be a summation of two types. But we think that this atypical presentation was not a simply superposition, but a result of the interaction between two pathogenic gene mutations.

To date several cases of double mutations associated with TGFBI gene have been described but the clinical phenotype of affected patients differs markedly from each other. Ha *et* $al^{[20]}$ observed a heterozygous P501T mutation of TGFBI gene



Figure 3 Sequence analysis of exon 4 and exon 12 of TGFBI gene in the family A: The exon 4 of proband exhibits a C > T heterozygous substitution at nucleotide position 417 and results in a arginine to cystine acid variation (R124C); B: A normal sequence of exon 4 of TGFBI encodes arginine in the codon 124; C: The exon 12 of proband exhibits a C > A heterozygous substitution at the 1637 position and results in a alanine to aspartic acid variation (A546D); D: A normal sequence of exon 12 is shown, which encodes alanine in codon 546.

and a homozygous Q118X mutation of MISI gene in same patient. The MISI gene was identified as responsible for the typical gelatinous drop like corneal dystrophy (GDLD) and usually in the autosomal recessive inheritance. It was reported that P501T mutation of the TGFBI gene could cause LCD $IIIA^{[10]}$. Ha *et al*^[20] finded that the patient with P501T and



Figure 4 The histopathological results of excised corneal stained with Congo red of the proband Light microscopy observed irregularly shaped and variably sized deposits in the anterior and middle stroma of the corneal tissue. A:10×; B:40×.

Q118X resembled GDLD penotype but not LCD. Dighiero et $al^{[21]}$ reported a French family affected with GCD and founded two heterozygous mutations in the TGFBI gene-R124L and Δ T125- Δ E126. These two mutations caused a variant of GCD that was intermediate in severity between the classical and superficial variant forms. Klintworth *et al*^[14] and Aldave *et al*^[22] presented a phenotypic variant of LCD associated with the A546D and P551Q missense changes in exon 12 of the TGFBI gene. The affected members had not only lattice-like corneal stromal deposits but also discrete, short, irregularly shaped opacities. Klintworth et al^[14] believed the P551Q allele could play a modifier role on the phenotype that associated with the A546D mutation. Yamada et al^[15] reported two cases with a double mutations in the TGFBI gene (R124H and N544S). The phenotype of the patients seemed to be a summation of the LCD and ACD. To sum up, these double mutations more or less produced phenotypic variations. It indicated that the interaction between these pathogenic mutations was exist and these alleles were modified by each other.

The modifier genes could influence the phenotypic variability in some monogenic disorders had been proven^[23]. And many factors such as environment, age, and sex may play an important role on expressivity and penetrance of the disease^[24]. However, the genetic interactions between mutated alleles and the modifier alleles or the effect of environmental factors are unclear. If those genetic interactions and modifier alleles can be confirmed, it may lead to a possible way to inhibit or delay the occurrence of genetic diseases.

In conclusion, our study observed a novel LCD pedigree which carried two pathogenic mutations (R124C and A546D) in the TGFBI gene. Its phenotypic features were obviously different from that reported LCD types. The result confirms that although the definite mutation is the most important genetic cause of disease, some different modifier alleles may influence the phenotype to some extent. Uncovering the interactions between them may lead to a possible way to inhibit the occurrence of the corneal dystrophy.

ACKNOWLEDGEMENTS

Our team sincerely thanks to all patients, their families and the volunteers for their participating in our research.

Foundations: Supported by the Ph.D. Programs Foundation of Heilongjiang Province (No.LBH-Q13126); the Research Foundation of the First Affiliated Hospital, Harbin Medical University (No.2011BS017).

Conflicts of Interest: Cao WP, None; **Yuan HG**, None; **Liu P**, None; **Li X**, None; **Hu Q**, None.

REFERENCES

1 Skonier J, Neubauer M, Madisen L, Bennett K, Plowman GD, Purchio AF. cDNA cloning and sequence analysis of beta ig-h3, a novel gene induced in a human adenocarcinoma cell line after treatment with transforming growth factor-beta. *DNA Cell Biol* 1992;11(7):511-522.

2 Munier FL, Korvatska E, Djemai A, Le Paslier D, Zografos L, Pescia G, Schorderet DF. Kerato-epithelin mutations in four 5q31-linked corneal dystrophies. *Nat Genet* 1997;15(3):247-251.

3 Hao XD, Zhang YY, Chen P, Li SX, Wang Y. Uncovering the profile of mutations of transforming growth factor beta-induced gene in Chinese corneal dystrophy patients. *Int J Ophthalmol* 2016;9(2):198-203.

4 Chae H, Kim M, Kim Y, Kim J, Kwon A, Choi H, Park J, Jang W, Lee YS, Park SH, Kim MS. Mutational spectrum of Korean patients with corneal dystrophy. *Clin Genet* 2016;89(6):678-689.

5 Cho KJ, Mok JW, Na KS, Rho CR, Byun YS, Hwang HS, Hwang KY, Joo CK. TGFBI gene mutations in a Korean population with corneal dystrophy. *Mol Vis* 2012;18:2012-2021.

6 Maury CP, Kere J, Tolvanen R, de la Chapelle A. Finnish hereditary amyloidosis is caused by a single nucleotide substitution in the gelsolin gene. *FEBS Lett* 1990;276(1-2):75-77.

7 de la Chapelle A, Kere J, Sack GH Jr, Tolvanen R, Maury CP. Familial amyloidosis, Finnish type: G654-a mutation of the gelsolin gene in Finnish families and an unrelated American family. *Genomics* 1992;13(3):898-901.

8 Tsujikawa M, Kurahashi H, Tanaka T, Okada M, Yamamoto S, Maeda N, Watanabe H, Inoue Y, Kiridoshi A, Matsumoto K, Ohashi Y, Kinoshita S, Shimomura Y, Nakamura Y, Tano Y. Homozygosity mapping of a gene responsible for gelatinous drop-like corneal dystrophy to chromosome 1p. *Am J Hum Genet* 1998;63(4):1073-1077.

9 Kim J, Lee KA, Kim EK, Lee HK. A Korean patient with lattice corneal dystrophy type IV with Leu527Arg mutation in the TGFBI gene. *Korean J Ophthalmol* 2014;28(1):83-85.

10 Yamamoto S, Okada M, Tsujikawa M, Shimomura Y, Nishida K, Inoue Y, Watanabe H, Maeda N, Kurahashi H, Kinoshita S, Nakamura Y, Tano Y. A kerato-epithelin (betaig-h3) mutation in lattice corneal dystrophy type IIIA. *Am J Hum Genet* 1998;62(3):719-722.

11 Jung JW, Kim SA, Kang EM, Kim TI, Cho HS, Kim EK. Lattice corneal dystrophy type IIIA with hyaline component from a novel A620P mutation and distinct surgical treatments. *Cornea* 2014;33(12):1324-1331. 12 Lai K, Reidy J, Bert B, Milman T. Spheroidal degeneration in H626R TGFBI variant lattice dystrophy: a multimodality analysis. *Cornea* 2014;33(7):726-732.

13 Costagliola C, Romano V, Cifariello F, Aceto F, Porcellini A. Lattice Corneal Dystrophy: a report of two cases in twin sisters due to 3 mutations (T1620C, C1416T, A1924G) in the TGFBI (BIGH3) gene. *Clin Ter* 2014;165(1):e73-e75.

14 Klintworth GK, Bao W, Afshari NA. Two mutations in the TGFBI (BIGH3) gene associated with lattice corneal dystrophy in an extensively studied family. *Invest Ophthalmol Vis Sci* 2004;45(5):1382-1388.

15 Yamada N, Kawamoto K, Morishige N, Chikama T, Nishida T, Nishioka M, Okayama N, Hinoda Y. Double mutation (R124H, N544S) of TGFBI in two sisters with combined expression of Avellino and lattice corneal dystrophies. *Mol Vis* 2009;15:974-979.

16 Afshari NA, Mullally JE, Afshari MA, Steinert RF, Adamis AP, Azar DT, Talamo JH, Dohlman CH, Dryja TP. Survey of patients with granular, lattice, avellino, and Reis-Bucklers corneal dystrophies for mutations in the BIGH3 and gelsolin genes. *Arch Ophthalmol* 2001;119(1):16-22.

17 Song JS, Lim DH, Chung ES, Chung TY, Ki CS. Mutation analysis of the TGFBI gene in consecutive Korean patients with corneal dystrophies. *Ann Lab Med* 2015;35(3):336-340.

18 Eifrig DE Jr, Afshari NA, Buchanan HW 4th, Bowling BL, Klintworth GK. Polymorphic corneal amyloidosis: a disorder due to a novel mutation in the transforming growth factor beta-induced (BIGH3) gene. *Ophthalmology* 2004;111(6):1108-1114.

19 Correa-Gomez V, Villalvazo-Cordero L, Zenteno JC. The TGFBI A546D mutation causes an atypical type of lattice corneal dystrophy. *Mol Vis* 2007;13:1695-1700.

20 Ha NT, Fujiki K, Hotta Y, Nakayasu K, Kanai A. Q118X mutation of M1S1 gene caused gelatinous drop-like corneal dystrophy: the P501T of BIGH3 gene found in a family with gelatinous drop-like corneal dystrophy. *Am J Ophthalmol* 2000;130(1):119-120.

21 Dighiero P, Drunat S, D'Hermies F, Renard G, Delpech M, Valleix S. A novel variant of granular corneal dystrophy caused by association of 2 mutation in the TGFBI gene-R124L and DeltaT125-DeltaE126. *Arch Ophthalmol* 2000;118(6):814-818.

22 Aldave AJ, Gutmark JG, Yellore VS, Affeldt JA, Meallet MA, Udar N, Rao NA, Small KW, Klintworth GK. Lattice corneal dystrophy associated with the Ala546Asp and Pro551Gln missense changes in the TGFBI gene. *Am J Ophthalmol* 2004;138(5):772-781.

23 Badano JL, Leitch CC, Ansley SJ, May-Simera H, Lawson S, Lewis RA, Beales PL, Dietz HC, Fisher S, Katsanis N. Dissection of epistasis in oligogenic Bardet-Biedl syndrome. *Nature* 2006;439(7074):326-330.

24 Cao W, Ge H, Cui X, Zhang L, Bai J, Fu S, Liu P. Reduced penetrance in familial Avellino corneal dystrophy associated with TGFBI mutations. *Mol Vis* 2009;15:70-75.