

Riboflavin/ultraviolet A-induced collagen cross-linking in rabbit corneal scar

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Received: 2018-02-10 Accepted: 2018-10-15

Abstract

• **AIM:** To evaluate the biomechanical stability of the corneal scar treating with riboflavin and ultraviolet A (UVA).

• **METHODS:** Totally 86 New Zealand rabbits were divided into control group (group A, $n=8$) and trauma groups [group B ($n=27$), group C ($n=24$) and group D ($n=27$)]. Then groups B, C and D were divided into three sub-groups according to the time points of sacrifice, *i.e.* groups Ba, Ca and Da (4wk, $n=8$); Bb, Cb and Db (6wk, $n=8$); Bc ($n=11$), Cc ($n=8$) and Dc (8wk, $n=11$). The right corneas of these 78 rabbits in the trauma groups were penetrated. Group B were only sutured. Group C were treated with corneal cross-linking (CXL) immediately after suturing. Group D were treated with CXL seven days after suturing. The corneal scar strips of $4.0 \times 10.0 \text{ mm}^2$ were cut and the stress and Young's modulus at 10% strain were evaluated. Samples from the three rabbits of group Bc and three of group Dc were used to measure the expression of alpha smooth muscle action (α -SMA).

• **RESULTS:** The mechanical strength of the corneal scar increased with time, and was strongest at 8wk after the injury. The ultimate stress of corneal scar (group D) were $2.17 \pm 0.52 \text{ MPa}$, $2.92 \pm 0.63 \text{ MPa}$, and $4.21 \pm 0.68 \text{ MPa}$ at 4wk, 6wk and 8wk, respectively; Young's modulus were $10.94 \pm 1.57 \text{ MPa}$, $11.16 \pm 2.50 \text{ MPa}$, and $13.36 \pm 2.10 \text{ MPa}$, which were higher than that of other groups except for normal control. The expression of α -SMA in group B and group D were 0.28 ± 0.11 and 0.65 ± 0.20 , respectively, and the difference was statistically significant ($P=0.048$).

• **CONCLUSION:** CXL with riboflavin/UVA at seven days after suturing improved the biomechanical properties of corneal scars most effectively in the present study.

• **KEYWORDS:** crosslinking; cornea; biomechanics; corneal penetrating injury; rabbit

DOI:10.18240/ijo.2019.01.07

Citation: Cai YH, Liu TX, Li HX. Riboflavin/ultraviolet A-induced collagen cross-linking in rabbit corneal scar. *Int J Ophthalmol* 2019;12(1):46-50

INTRODUCTION

The cornea is the transparent tissue covering the front of the eye. The penetrating corneal trauma is very common due to directly contacting the external environment^[1]. Epithelial-stromal injury of the cornea initiates a complex stromal response that can lead to the formation of corneal scar^[2-3]. The scar leads to opacity of the cornea and weakened biomechanical properties. When intraocular pressure increases continuously, corneal scar could develop to corneal staphyloma. Corneal scar tissue's impact on biomechanical properties of the cornea has been documented in an experiment report^[4].

The biomechanics of collagen depends on the collagen density and the covalent cross-linking between collagen molecules^[5]. Therefore, cross-linking plays an important role in the strength of collagen tissue. Over the past decade, a technique to generate corneal collagen cross-linking (CXL) was invented by Seiler^[6]. CXL induced by riboflavin/ultraviolet A (UVA) has a stiffening effect that increases resistance of the collagen against proteolytic enzymes, and increases the collagen diameter which could improve the biomechanics of the cornea^[7-12]. CXL was originally designed to improve the corneal biomechanics and prevent further development of keratoconus^[13]. CXL was also used for the treatment of corneal ulcers, which healed with scar tissue and no complications^[14-15]. CXL has also been reported successfully in preventing the progress of keratectasia after refractive surgery^[16]. To the best of our knowledge, there was no report about CXL in corneal scar tissue which induced by penetrating corneal trauma. The purpose of the present study is to evaluate the biomechanics of corneal scar which treated with riboflavin/UVA.

MATERIALS AND METHODS

Ethical Approval The experiments and all animal procedures were approved by the ethics committee of Zunyi Medical College, and followed guidelines of the Association for Research in Vision and Ophthalmology Statement for the Use of Animals in Ophthalmic and Vision Research.

Animals Eighty-six healthy rabbits were selected and divided into control group (group A, $n=8$) and trauma groups

[group B ($n=27$), group C ($n=24$) and group D ($n=27$)]. The right corneas of these 78 rabbits in the trauma groups were penetrated using blades. The corneas in group B were treated only with sutures. The corneas in group C were treated with riboflavin/UVA immediately after suturing. The corneas in group D were treated with riboflavin/UVA seven days after suturing. Then groups B, C and D were further divided into subgroups according to the time points of sacrifice, *i.e.* groups Ba (4wk, $n=8$), Bb (6wk, $n=8$) and Bc (8wk, $n=11$); groups Ca (4wk, $n=8$), Cb (6wk, $n=8$) and Cc (8wk, $n=8$); groups Da (4wk, $n=8$), Db (6wk, $n=8$) and Dc (8wk, $n=11$). Samples from the three rabbits of group Bc and three of group Dc were used to measure the expression of alpha smooth muscle action (α -SMA).

Corneal Penetrating Injury Model Rabbits were anesthetized by intravenous injection of 2% sodium pentobarbital solution (Beijing Pubos Biotechnology Co., Ltd. Beijing, China) and then a full-thickness incision about 6 mm length was cut in the vertical direction around the centre of cornea. The incisions were sutured intermittently with four stitches using 10-0 nylon suture.

Riboflavin/UVA Cross-linking According to Wollensak's method^[17], 30min before the irradiation, 0.1% riboflavin photosensitizer solution (10 mg riboflavin-5-phosphate in 10 mL 20% dextran-T-500; Beijing Solarbio Science & Technology Co., Ltd., Beijing, China) was dropped on the target cornea. UVA irradiation (370 nm) was applied using a single UVA diodes (Zhuhai Tianhui Electronic Co., Zhuhai, China) with an irradiance of 3 mW/cm² for 30min. During the irradiation, the riboflavin was instilled on the cornea every 3min. Illumination intensity was monitored using a calibrated UV light meter (UV-340A, 290-390 nm, Taiwan, China).

Stress-strain Test At the time points of sacrifice, the rabbits were killed by air embolism. A corneal strip (4.0 mm wide, 10.0 mm in length) was cut using a double-blade scalpel. The scar must be placed in the middle of the strip. The corneal strips were preserved less than 24h at 4°C in a moist chamber. Then the corneal scar thickness was measured using a digimatic calliper [Sanling group (H.K.) Ltd., H.K., China] and a stress-strain test was performed.

The stress-strain tests of the corneal strips were performed using CMT6104 (Meitesi industry system Co. Ltd., Minnesota, USA). The strips were subjected to a stress level of 0.02 MPa and 5 cycles in total of deformation-load testing. Strain was increased linearly at a velocity of 1.5 mm/min, and stress was measured up to scar fracture.

Western Blot Corneal tissues from the three rabbits of group Bc and three of group Dc were excised and homogenized in the immunoprecipitation assay (RIPA) lysis buffer. Proteins in whole lysate (10 μ g protein per sample) were electrophoresed

Table 1 Ultimate stress and modulus at 10% strain mean \pm SD

Groups	Treatment	Stress (MPa)	Modulus (MPa)
A	control	5.71 \pm 1.29	16.37 \pm 4.06
Ba	suture-4w	1.37 \pm 0.62	4.64 \pm 1.69
Bb	suture-6w	2.21 \pm 0.30	7.75 \pm 0.82
Bc	suture-8w	3.14 \pm 0.74	9.02 \pm 0.87
Ca	CXL (immediate)-4w	1.87 \pm 0.56	9.67 \pm 2.42
Cb	CXL (immediate)-6w	2.83 \pm 0.54	10.67 \pm 2.07
Cc		3.26 \pm 0.83	11.08 \pm 1.61
Da	CXL (immediate)-8w	2.17 \pm 0.52	10.94 \pm 1.57
Db	CXL (7d)-4w	2.92 \pm 0.63	11.16 \pm 2.50
Dc	CXL (7d)-6w	4.21 \pm 0.68 ^a	13.36 \pm 2.10 ^a
F	CXL (7d)-8w	23.94	15.69
P		0.00	0.00

^a $P<0.05$.

by 7.5% SDS-PAGE and then electroblotted onto nitrocellulose membranes which were blocked in 5% skim milk. Subsequently, the membrane was incubated overnight with primary antibodies directed against actin and α -SMA antibody (1:5000, Beijing Boersen Biotechnology Co., Ltd., Beijing, China) at 4°C. The membranes were washed using tris-buffered saline with Tween 20 (TBST, Cell Signaling Technology) and incubated with a horseradish peroxidase-conjugated secondary antibody for 60min at room temperature (β -actin 1:5000; BoAoSeng, Beijing, China). Chemiluminescence assays were processed using a peroxidase substrate. The immunoblot signal was detected and analyzed using Image J software.

Statistical Analysis The ultimate stress and Young's modulus data at 10% strain were expressed as mean \pm standard deviation and analysed by the one-way ANOVA (version 17.0, SPSS Inc, Chicago, IL, USA). Normality for continued variables in groups was analyzed using the one-way ANOVA. A value of $P<0.05$ was considered statistically significant.

RESULTS

The mean thickness of the corneal scar strips was 560 \pm 84 μ m. There was statistically significant difference among all groups ($P<0.05$). Group A was 398 \pm 7 μ m; groups Ba, Bb and Bc were 640 \pm 62 μ m, 609 \pm 63 μ m and 579 \pm 63 μ m, respectively; groups Ca, Cb and Cc were 604 \pm 69 μ m, 581 \pm 76 μ m and 537 \pm 26 μ m, respectively; groups Da, Db and Dc were 603 \pm 82 μ m, 533 \pm 44 μ m and 524 \pm 19 μ m, respectively.

When compared the value of the ultimate stress and Young's modulus at 10% strain, corneal strips in CXL groups had higher values than group B, but still lower than the control group ($P<0.05$). Group Dc had a significant higher value of the stress and modulus than those in other trauma groups (groups B, C, Da and Db; $P<0.05$; Table 1).

The stress-strain values increased exponentially in all study groups. The ultimate stress and Young's modulus of the corneal strips increased when the time after the injury getting longer

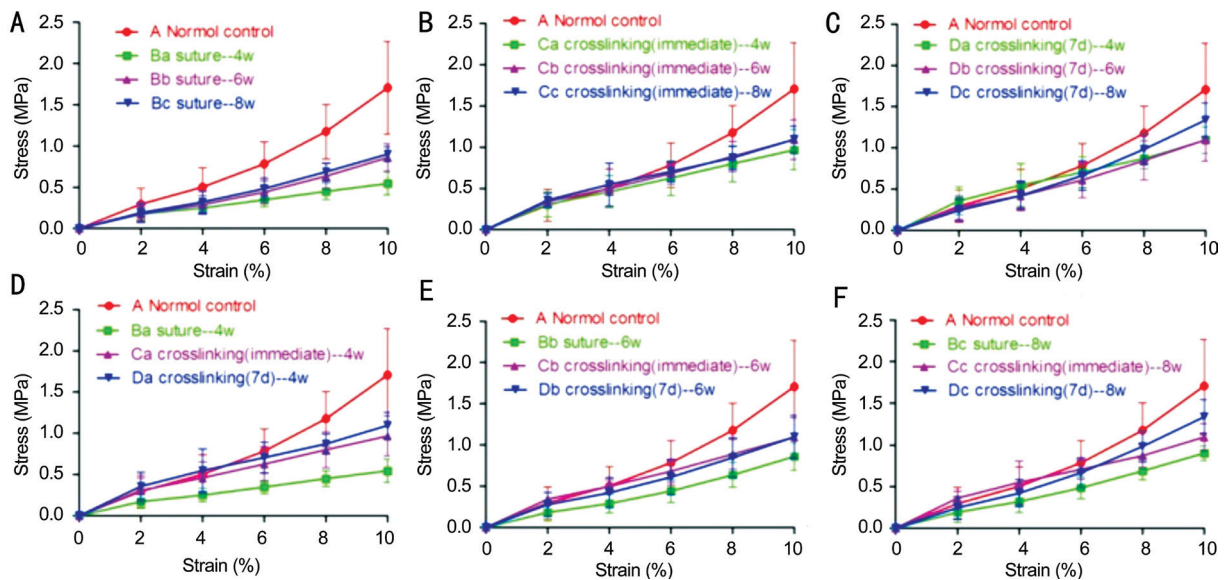


Figure 1 The stress-strain values showed an exponential increase in all groups. The ultimate stress and young's modulus increased with time (A-C). The largest values were observed at 8wk ($P<0.05$). The values from the CXL groups were lower than that in the control group ($P=0.00$). The stress of corneal scar in group B was lower than that of group D ($P<0.05$, D-F).

(Figure 1A-1C). The highest value in the trauma groups was obtained in group Dc ($P<0.05$). However, the value of group Dc was still lower than that of the control group ($P<0.01$). The stress values of corneal strips in group D were significantly higher than those in group B ($P<0.05$; Figure 1D-1F).

α -SMA was measured by Western blot in groups Bc and Dc. The relative expression of α -SMA were 0.28 ± 0.11 and 0.65 ± 0.20 , respectively. The difference of the above values was statistically significant ($t=-2.48$, $P=0.048$; Figure 2).

DISCUSSION

The cornea is a viscoelastic tissue that is composed of a large number of collagen fibers. The polar region of a collagen molecule cross-links the non-polar region of another collagen molecule *via* covalent and electrostatic attraction. Thus, the collagen fibers have very high tenacity. Because of its high tenacity and strong tensile resistance, collagen fibers play an important role in maintaining corneal tension^[18]. When the collagen structure is affected by external stimuli, its chemical and physical properties would change, such as elasticity and stiffness. The corneal wound healing response usually contributes to a restore of normal stromal structure and function^[19-20]. During the repair process after injury, fibroblasts underwent the migration and proliferation, and part of them would transform into myofibroblasts^[21-22]. The biomechanical properties of the corneal scar after the excimer laser *in situ* keratomileusis surgery was found weak and easily broken in bruises. Clinically, corneal scar is the prominent location of corneal staphyloma, and it also indicates that the resistance of scar tissue to intraocular pressure is decreased. To protect the corneal endothelium, lens and retina, the minimum corneal thickness for performing CXL is $400\ \mu\text{m}$ ^[23-25]. In the present

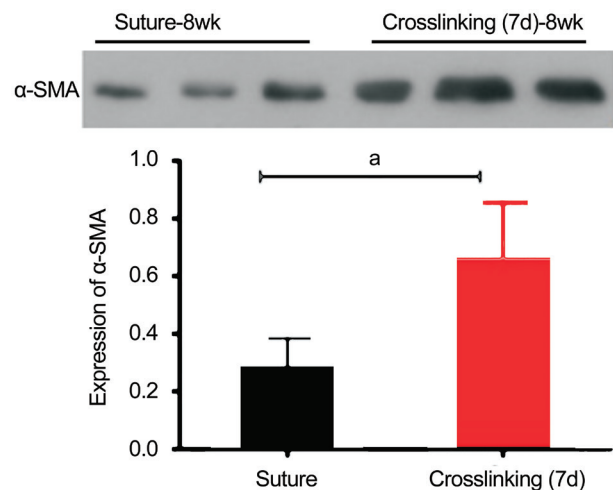


Figure 2 Relative expression of α -SMA evaluated by Western blotting. Value in group Dc was significantly higher than that in group Bc. ^a $P<0.05$.

study, the mean thickness of corneal scars was $560\pm 84\ \mu\text{m}$. Our experiment was safe to the rabbits.

The commonly used index to evaluate the biomechanical properties of a viscoelastic tissue is Young's modulus, which is the ratio of stress to strain^[26-27]. There was a report that after corneal penetrating injury, the tension of the rabbit cornea increased with the prolongation of time and reaches maximum at 6wk^[28]. However, in the present study, the corneal tensile strength gradually increased and reached a highest value at 8wk which was the final time point of the study. The reason of a longer restore time in our study than that in the literature may due to a larger lesion we made than the lesion size reported in the other study. Because when the incision was larger, the stability of the cornea was worse, and the recovery time of the tissue repair was increased. The results of the present

study demonstrated that CXL increased the biomechanical characteristics of the scar formed in the penetrating corneal injury. CXL could be used to remove sutures earlier when a corneal penetrating injury occurs. Our data showed that the best effect of improving biomechanics of a corneal scar could be induced by CXL at seven days after trauma. We speculated that it may be due to the activation and transdifferentiation of fibroblasts to myofibroblasts, and large amount of collagen was synthesized.

Myofibroblasts express α -SMA which is a cytoskeletal protein^[29] and it can make myofibroblasts contractile^[30]. The corneal tissue does not express α -SMA. The expression of α -SMA can be used to determine whether the myofibroblasts appear after the corneal trauma^[31-32]. There was study report that the epithelial cells proliferated significantly and the number of fibroblasts increased after photorefractive keratectomy, and the expression of α -SMA was obvious^[33]. α -SMA connects with extracellular collagen and fibronectin via integrin α 2b1 and α 5b1, enhances the ability of wound tissue to resist external tension^[34]. α -SMA has been reported that has impact on biomechanics of the sclera^[35]. The expression of α -SMA was observed at one week after corneal trauma occurred. CXL may increase the expression of α -SMA by affecting myofibroblasts. Our Western blotting tests showed that expression of α -SMA was significantly increased in the group performing CXL at seven days after corneal lesion suturing. It documented that CXL induced by riboflavin/UVA increased α -SMA and enhanced the biomechanical properties of corneal scar tissue. Further studies are needed to confirm the long term effect of CXL with riboflavin/UVA, which could be a potential therapy to prevent the progression of corneal staphyloma.

ACKNOWLEDGEMENTS

Foundations: Supported by the National Natural Science Foundation of China (No.81660169); the Science and Technology Foundation of Zunyi [No.(2014)94].

Conflicts of Interest: Cai YH, None; Liu TX, None; Li HX, None.

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