

Leptin's concentration in tears and dry eye: a clinical observational study

Ran Hao, Yan Liu, Xue-Min Li

Department of Ophthalmology, Peking University Third Hospital, Beijing 100191, China

Correspondence to: Xue-Min Li. Peking University Third Hospital, No. 49, North Garden Street, Beijing 100191, China. 13911254862@163.com

Received: 2019-12-23 Accepted: 2020-07-27

Abstract

• **AIM:** To investigate the concentration of leptin in tears and its correlation with dry eye symptoms and signs.

• **METHODS:** The study enrolled individuals ($n=39$) responding to an advertising or dry eye patients ($n=58$) from the Ophthalmology Department. Tear samples were collected for leptin concentration measuring. Ocular Surface Disease Index (OSDI), tear meniscus height (TMH), tear break up time (TBUT), cornea fluorescein staining, Schirmer test (ST) and impression cytology (IC) were assessed. Leptin concentration in tears of dry eye patients and healthy controls, and its correlation with clinical features of dry eye disease (DED) were analyzed.

• **RESULTS:** Age, body mass index (BMI), OSDI scores and cornea fluorescein staining scores showed a negative correlation with leptin concentration in tears ($r=-0.340$, $P=0.001$; $r=-0.332$, $P=0.001$; $r=-0.258$, $P=0.011$; $r=-0.424$, $P<0.001$, respectively). ST showed positive correlation with leptin concentration in tears ($r=0.206$, $P=0.045$). No significant difference was observed in leptin concentration between dry eye patients and controls ($P=0.682$). Multivariate linear regression analysis revealed that dry eye, OSDI, corneal fluorescein staining scores and ST correlated with leptin concentration in tears.

• **CONCLUSION:** This is the first study measuring leptin concentration in tears. The correlation between leptin concentration and DED symptoms and signs reveal that leptin level correlated with the dry eye, potentially contributing to repair of ocular damage and dry eye improvement.

• **KEYWORDS:** dry eye disease; leptin; ocular surface inflammation; wound healing

DOI:10.18240/ijo.2021.01.12

Citation: Hao R, Liu Y, Li XM. Leptin's concentration in tears and dry eye: a clinical observational study. *Int J Ophthalmol* 2021;14(1):83-88

INTRODUCTION

Dry eye disease (DED) is a common ocular disorder characterized by ocular surface discomfort and visual impairment^[1]. Reduced quality and quantity of tears due to reduced secretion or excessive evaporation, causes increased osmolality of the tear film and promotes ocular surface inflammation^[1-2].

Increasing evidence demonstrates that ocular surface inflammation is responsible for the dry eye-associated symptoms and signs^[1-8]. Also, research indicates that long-term tear film anomalies decrease anti-inflammatory components, such as lactoferrin, and increases pro-inflammatory cytokines in tears, such as interleukin-1 β (IL-1 β), tumor necrosis factor- α (TNF- α) and protease matrix metalloproteinase-9 (MMP-9). These inflammatory mediators appear to activate the mitogen-activated protein kinases (MAPKs) pathway and initiate inflammatory cascades on the ocular surface, which stimulates inflammatory cytokines and proteases expression^[8]. The inflammatory cascades lead to increasing expression of immune activation and cell adhesion molecules such as intercellular cell adhesion molecule-1 (ICAM-1) and human leukocyte antigen-DR (HLA-DR), recruiting more inflammatory cells and aggravating inflammation^[8-10]. Molina-Leyva *et al*^[11] found that the levels of inflammatory cytokines such as IL-1, IL-2, IL-6, IL-8 and TNF in tears and in the conjunctival epithelium of dry eye patients, were higher than in normal patients, in direct proportion to the severity of DED. Leptin is a unique member of the cytokine family that is a product of the Ob gene^[12-15]. Leptin plays a role in inhibiting food intake, adjusting energy consumption, regulating reproduction and development, promoting angiogenesis and cell proliferation as well as regulating immune responses and inflammatory processes^[12,15-21]. Leptin has a similar structure to Class I cytokines and may function in a similar way to IL-6, leukemia inhibitory factor (LIF) and granulocyte colony-stimulating factor (G-CSF), among others^[12,14]. Recent studies demonstrate that leptin acts as a pro-inflammatory cytokine and regulates inflammatory cytokines secretion in varying patterns^[13,16,19]. Different inflammatory stimuli have been found to regulate leptin expression^[12,15,19]. Indeed, the increased level of circulating leptin in acute inflammation may support initial pro-inflammatory responses^[19].

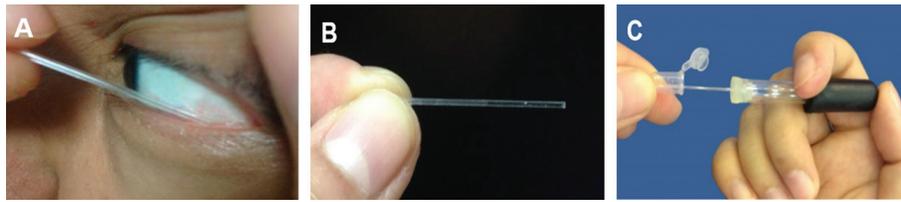


Figure 1 Tear collection A: A plastic head was used to squeeze tear into a capillary tube from the lower tear meniscus; B: Tears inside the capillary tube; C: Tears were collected from the capillary tube and placed in EP tubes.

Increasing evidence indicates that leptin correlates with the development of ocular inflammation and DED^[16]. Serum leptin significantly increases in patients with infectious keratitis and promote pterygium progression in animal models^[16]. Leptin can also stimulate the formation of corneal, retinal and choroidal neovascularization in animal models^[17-21]. Based on this reports, we aim to study whether leptin is related to DED-associated ocular surface inflammation.

The purpose of this study is to measure the concentration of leptin in the tears and evaluate its correlation with DED symptoms and signs.

SUBJECTS AND METHODS

Ethical Approval The study was performed in accordance with the Declaration of Helsinki and approved by the Human Research and Ethics Committee of the hospital where the study was conducted. Written informed consent was obtained from all study subjects.

Study Design and Enrollment This observational study includes fifty-eight dry eye patients (58 eyes) from the Department of Ophthalmology, and healthy volunteers (39 eyes) that responded to the advertising without DED signs or symptoms or other ocular problems, enrolled from April 2014 to January 2015.

DED patients must had at least 1 symptom (foreign body sensation or eye irritation or light sensitivity or grittiness) and 1 sign [Schirmer test (ST) ≤ 5 mm/5min or corneal fluorescein staining score ≥ 4] to be included in the study. The right eye of each participant was enrolled in the study. Patients were excluded from the study if they were pregnant women, children, any patient with acute inflammation and serious systemic disease, using systemic steroids, anti-inflammatory or antimetabolites. All the examinations were performed by the same experienced clinician, who was masked to group aleatorization of patients.

Symptom Evaluation Symptomatic evaluation with the ocular surface disease index (OSDI) preceded clinical ocular surface examination. This questionnaire is comprised of 12 questions and evaluates the frequency of symptoms over the preceding week. OSDI is divided into three parts: eye discomfort, visual function impairment and susceptibility to environmental factors, with scores that range from 0 to 100^[22].

Tear Collection and Leptin Concentration Measurement

Non-irritating tear collection was conducted using 5 μ L capillary pipets without any anaesthetic (Figure 1). Patients did not receive eye drops prior to collection. A plastic head was used to squeeze tears into 0.2 mL EP tubes and tears were immediately placed at -80°C for storage. The level of leptin in the specimen was measured using a double antibody sandwich assay with the human leptin enzyme-linked immunoassay (ELISA) kit (Abcam, United Kingdom).

Clinical Evaluation The following diagnostic tests were also performed: tear meniscus height (TMH), tear film break up time (TBUT), corneal fluorescein staining, ST and impression cytology (IC). We used a micro-ruler to measure central TMH at the lower eyelid under slit-lamp (10 \times magnification). TBUT was observed under cobalt blue illumination. Corneal staining with sodium fluorescein was graded using the Van Bijsterveld scale. The cornea was divided into 4 quadrants, and scored as follows, 0: no staining; 1: 1-6 spots; 2: 7-30 spots; 3: >30 spots^[23]. The number of spots was counted in each quadrant, to obtain a total score. The IC samples were obtained from similar locations (temporal palpebral conjunctiva). Five areas were randomly selected to count goblet cell number on IC image, and the mean was calculated accordingly under a microscope ($\times 400$) and graded by Nelson's scale as follows^[24], 0: small and round cells and large nuclei, with a nuclear cytoplasmic ratio of 1:2, goblet cells >500 cells/ mm^2 , PAS positive staining; 1: slightly larger cells and smaller nuclei, with a nuclear cytoplasmic ratio of 1:3, goblet cells are decreased to 350-500 cells/ mm^2 , but maintain PAS positive staining; 2: larger and polygonal cells and small nuclei, with nucleocyto-plasmic ratio of 1:4-1:5, goblet cells 100-350 cells/ mm^2 , poorly-defined borders with less intensively PAS-positive; 3: more larger and polygonal cells and small and pyknotic nuclei, with a nuclear cytoplasmic ratio of $>1:6$, goblet cells <100 cells/ mm^2 .

Statistical Analysis SPSS 19.0 (IBM, Armonk, NY, USA) statistical software was used for statistical analysis. Descriptive parameters were expressed as the number of patients (%), means \pm standard deviation (SD) or median (Q1, Q3) based on the distribution and the homogeneity of variance of the data. Independent *t*-test was used to compare age, body mass index (BMI) and leptin concentration in tears between the two groups. Mann-Whitney *U* test was used to compare OSDI,

Demographics	Dry eye group (n=58)	Control group (n=39)	median (Q1, Q3) P
Gender			0.570
Male, n (%)	18 (31.03)	14 (35.90)	
Female, n (%)	40 (68.97)	25 (64.10)	
Age (y), mean±SD	56.16±19.39	52.23±22.33	0.409
BMI (kg/m ²), mean±SD	22.59±2.63	23.13±4.01	0.892
Eye discomfort score	47.5 (35, 60)	5 (0, 25)	<0.001 ^a
Visual impairment score	25 (0, 50)	0 (0, 25)	0.001 ^a
Susceptibility to environment score	16.67 (0, 50)	0 (0, 25)	0.057
OSDI overall score	34.17 (24.43, 52.78)	9.09 (0, 20.83)	<0.001 ^a
TMH (mm)	0.15 (0.1, 0.2)	0.2 (0.2, 0.3)	<0.001 ^a
TBUT (s)	3 (2.38, 4)	5 (3, 7)	<0.001 ^a
Corneal fluorescein staining (score)	2 (0, 5)	0 (0, 3)	0.042 ^a
ST (mm)	8 (5, 15)	15 (8, 22)	0.004 ^a
Goblet cells density(/mm ²)	78.25 (50.08, 136.16)	134.59 (82.95, 198.75)	0.003 ^a
Nelson's grade	2 (2, 3)	1.25 (1.25, 2)	<0.001 ^a

BMI: Body mass index; OSDI: Ocular surface disease index; TMH: Tear meniscus height; TBUT: Tear film break up time; ST: Schirmer test. ^aP<0.05.

TMH, TBUT, corneal fluorescein staining, ST and IC between the two groups. The correlation between leptin concentration in tears and clinical symptoms/signs was analyzed using Spearman's correlation coefficients. The effects of dry eye and clinical symptoms/signs on leptin concentration in tears were assessed by multivariate linear regression. Chi-square test was used to compare the gender composition. Results were considered significant at P<0.05.

RESULTS

A total of 97 eyes from 97 individuals (aged 54.58±20.60y, 65 females) were enrolled in the study, including 58 dry eye patients (aged 56.16±19.39y, 40 females) and 39 healthy controls (aged 52.23±22.33y, 25 females). No significant differences in gender, age and BMI were observed between groups (P=0.570, P=0.409, and P=0.892, respectively). Demographics are shown in Table 1.

Eye discomfort, visual impairment and total scores in dry eye patients were significantly higher compared to controls, with a significant difference between groups (P≤0.001). The sensitivity to the environment scores was numerically higher in the dry eye group but there was no significant difference between groups (P=0.057). The results are showed in Table 1 and Figure 2. TMH, TBUT and ST were significantly lower in dry eye patients and corneal fluorescein staining was significantly lower in the control group (P<0.001, P<0.001, P=0.004, and P=0.042, respectively; Table 1 and Figure 3).

A total of 86 IC images were obtained, 53 from the dry eye group and 33 from the control group. Goblet cell density in the dry eye group was 78.25 (50.08, 136.16/mm²), and in the control group was 134.59 (82.95, 198.75/mm²). Goblet cell

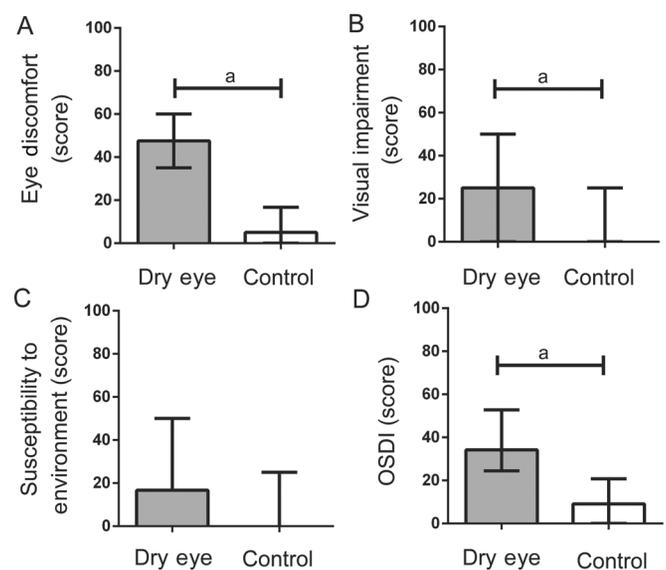


Figure 2 The scores of each OSDI subscales and overall score from dry eye patients and controls A: Eye discomfort; B: Visual impairment; C: Susceptibility to environment; D: Overall OSDI score. ^aP<0.05.

density in the control group was significantly higher than in the dry eye group (P=0.003; Figure 4A-4E). There was a statistically significant difference in Nelson's classification (Figure 4F) between the two groups (P<0.001).

Leptin concentration in tears was 5.04±1.08 µg/L in the dry eye group, and 5.14±1.12 µg/L in the control group. No significant difference was observed between groups (P=0.682). There was a significant negative correlation between leptin concentration and age (r=-0.340, P=0.001), BMI (r=-0.332, P=0.001), OSDI (r=-0.258, P=0.011) and corneal fluorescein staining

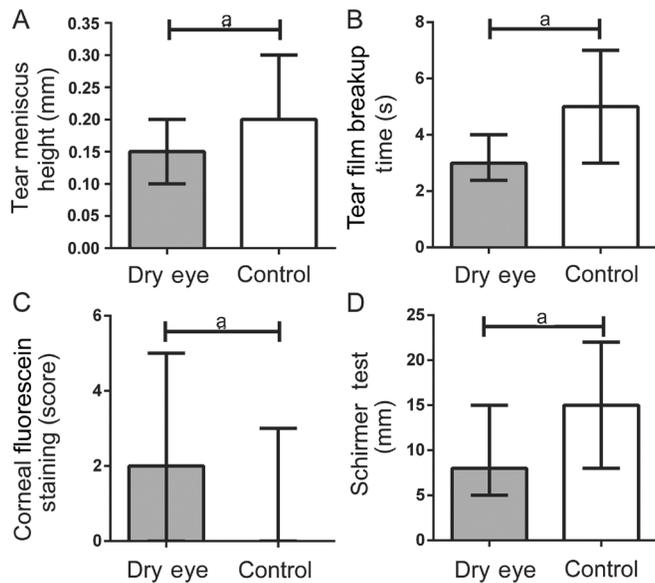


Figure 3 Results of dry eye signs in both groups A: Tear meniscus height; B: Tear break up time; C: Corneal fluorescein staining; D: ST. ^a*P*<0.05.

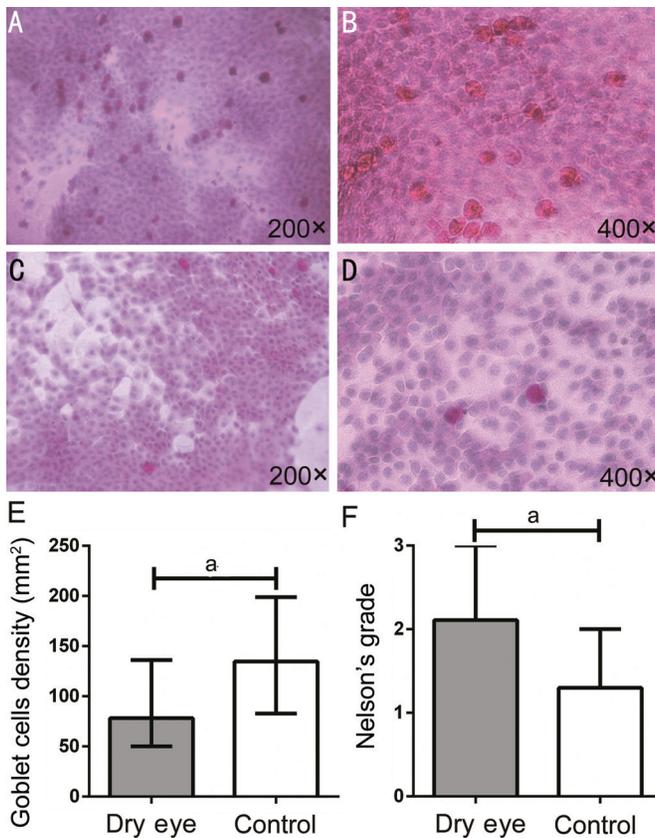


Figure 4 IC from dry eye patients and controls A: Normal IC (200×); B: Normal IC (400×); C: IC of dry eye patients (200×); D: IC of dry eye patients (400×); E: Goblet cell density; F: Nelson's grade. The goblet cells are plump and oval with a PAS-positive staining cytoplasm. ^a*P*<0.05.

scores ($r=-0.424$, $P<0.001$). Leptin concentration in tears was positively correlated with ST ($r=0.206$, $P=0.045$). However, the TMH, TBUT and goblet cell density were not significantly correlated with leptin concentration. Multivariate linear

Table 2 Multivariate linear regression models: correlation between independent variables and leptin concentration in tears

Items	β	<i>P</i>
Dry eye	0.599	<0.021 ^a
OSDI	-0.015	0.007 ^a
Corneal fluorescein staining scores	-0.136	<0.001 ^a
ST	0.041	0.001 ^a

OSDI: Ocular surface disease index; ST: Schirmer test; ^a*P*<0.05.

regression analysis revealed that 4 variables were associated with leptin concentration in tears (Table 2). We found the linear equation best fitting the data to be $Y=4.874+0.599X_1-0.015X_2-0.136X_3+0.041X_4$ ($r^2=0.252$, $P<0.05$), where *Y* represents the leptin concentration in tears and X_1 , X_2 , X_3 , X_4 , represent dry eye, OSDI, corneal fluorescein staining scores and ST, respectively. The strongest predictor was dry eye ($\beta=0.599$).

DISCUSSION

To the best of our knowledge, this is the first study measuring leptin concentration in tears and analyzing the correlation between dry eye symptoms and signs with leptin concentration, to explore the role of leptin in the development of dry eye.

In this study, we found no significant difference in leptin concentration ($P=0.682$), in tears of patients with dry eye and controls, which may be attributable to the low volume of tears collected from DED patients. Serum leptin concentration of normal-weight subjects has been positively correlated with absolute fat mass, percentage body fat and BMI^[25]. Gender differences also correlate with serum leptin levels in adults. When total body fat is similar, female serum leptin level is twice as high compared to male^[26], and the concentration of leptin in serum in pregnant women is higher than that of non-pregnant women^[27]. In this study, we found no significant differences in gender, age and BMI between the two groups, excluding an influence of demographic features on leptin levels. Also, we found that leptin concentration in the tears was negatively correlated with age, consistent with the change of leptin concentration in serum with age^[28]. Interestingly, leptin concentration in the tears was negatively associated with BMI, and it was different from serum leptin concentration^[29]. This may be due to a low number of samples and reduced tears volume.

A significant difference in eye discomfort, visual impairment and total scores was observed between groups, but no statistically significant differences were observed on the sensitivity to environmental factors. This suggests that patients with dry eye may have certain resistance to environmental stimuli. This may be due to a certain protection and injury restore mechanism of the ocular surface that enables patients with serious ocular discomfort and visual disorders to defend from external adverse stimuli.

Leptin can regulate the secretion of several cytokines secretion and different inflammatory stimuli could also regulate leptin expression and circulating leptin levels^[13,16,19]. Previous studies have found that a long term instability of tear film in dry eye patients led to ocular surface inflammation, which was noted to be highly correlated with dry eye symptoms, as well as with a number of clinical features^[1-3]. Leptin, as an inflammatory factor, also accumulates on the ocular surface; increased leptin concentration may accelerate mitosis in cells that surround an area of ocular surface damage, thereby regulating and accelerating the process of corneal epithelial cells regeneration^[30-33], promoting ocular fibroblast proliferation and collagen synthesis^[33], and stimulating angiogenesis. These processes help to repair ocular damage, alleviating dry eye symptoms and signs. Our findings, that increased leptin concentration in tears promotes symptom alleviation and improves corneal staining scores and tears secretion, are consistent with these studies.

Dry eye severity was associated with decreased goblet cell density, but the correlation of leptin concentration in tears with conjunctival goblet cell density failed to reach statistical significance. This may be due to the fact that dry eye symptoms and signs appear earlier than pathological changes like decreased goblet cell density^[1]. Increased leptin concentration could induce ocular injury repair, but cell morphological pathological changes observed in IC caused by the long-term ocular surface damage may more time to recover.

Our study has some limitations. This is a cross-sectional study which cannot determine an exact correlation between leptin concentration in tears and DED appearance. Further longitudinal studies are required to confirm this. A larger sample size is needed to investigate the physiopathology of leptin concentration in tears and dry eye severity.

In conclusion, this is the first study measuring leptin concentration in tears. No significant difference was observed in the tears leptin concentration between dry eye patients and controls. The correlation between leptin concentration and DED symptoms and signs probably revealed that leptin level is associated with the DED. Finally, leptin levels potentially contributed to ocular surface repair and DED alleviation. To further explain the findings of this study, investigations on the mechanism of increased leptin concentration in tears caused by dry eye and leptin effects on the ocular surface, are warranted.

ACKNOWLEDGEMENTS

Conflicts of Interest: Hao R, None; Liu Y, None; Li XM, None.

REFERENCES

1 Willcox MDP, Argüeso P, Georgiev GA, Holopainen JM, Laurie GW, Millar TJ, Papas EB, Rolland JP, Schmidt TA, Stahl U, Suarez T, Subbaraman LN, Uçakhan OÖ, Jones L. TFOS DEWS II tear film report. *Ocular Surf* 2017;15(3):366-403.

- 2 Clayton JA. Dry eye. *N Engl J Med* 2018;378(23):2212-2223.
- 3 Pflugfelder SC, de Paiva CS. The pathophysiology of dry eye disease: what we know and future directions for research. *Ophthalmology* 2017;124(11):S4-S13.
- 4 Martin E, Oliver KM, Pearce EI, Tomlinson A, Simmons P, Hagan S. Effect of tear supplements on signs, symptoms and inflammatory markers in dry eye. *Cytokine* 2018;105:37-44.
- 5 Rouen PA, White ML. Dry eye disease: prevalence, assessment, and management. *Home Healthc* 2018;36(2):74-83.
- 6 Satitpitakul V, Kheirkhah A, Crnej A, Hamrah P, Dana R. Determinants of ocular pain severity in patients with dry eye disease. *Am J Ophthalmol* 2017;179:198-204.
- 7 Galor A, Moein HR, Lee C, Rodriguez A, Felix ER, Sarantopoulos KD, Levitt RC. Neuropathic pain and dry eye. *Ocular Surf* 2018;16(1):31-44.
- 8 Luo LH, Li DQ, Doshi A, Farley W, Corrales RM, Pflugfelder SC. Experimental dry eye stimulates production of inflammatory cytokines and MMP-9 and activates MAPK signaling pathways on the ocular surface. *Investig Ophthalmol Vis Sci* 2004;45(12):4293-4301.
- 9 Rolando M, Barabino S, Mingari C, Moretti S, Giuffrida S, Calabria G. Distribution of conjunctival HLA-DR expression and the pathogenesis of damage in early dry eyes. *Cornea* 2005;24(8):951-954.
- 10 Liu RJ, Gao CF, Chen HJ, Li YX, Jin Y, Qi H. Analysis of Th17-associated cytokines and clinical correlations in patients with dry eye disease. *PLoS One* 2017;12(4):e0173301.
- 11 Molina-Leyva I, Molina-Leyva A, Bueno-Cavanillas A. Efficacy of nutritional supplementation with omega-3 and omega-6 fatty acids in dry eye syndrome: a systematic review of randomized clinical trials. *Acta Ophthalmol* 2017;95(8):e677-e685.
- 12 Zhang Y, Chua S Jr. Leptin function and regulation. *Compr Physiol* 2017;8(1):351-369.
- 13 Modan-Moses D, Kanety H, Dagan O, Ehrlich S, Lotan D, Pariente C, Novikov I, Paret G. Leptin and the post-operative inflammatory response. More insights into the correlation with the clinical course and glucocorticoid administration. *J Endocrinol Invest* 2010;33(10):701-706.
- 14 Madej T, Boguski MS, Bryant SH. Threading analysis suggests that the obese gene product may be a helical cytokine. *FEBS Lett* 1995;373(1):13-18.
- 15 Lanier V, Gillespie C, Leffers M, Daley-Brown D, Milner J, Lipsey C, Webb N, Anderson LM, Newman G, Waltenberger J, Gonzalez-Perez RR. Leptin-induced transphosphorylation of vascular endothelial growth factor receptor increases Notch and stimulates endothelial cell angiogenic transformation. *Int J Biochem Cell Biol* 2016;79:139-150.
- 16 Aydemir O, Naziroğlu M, Colakoğlu N, Yılmaz T, Kükner A, Kükner AS. Leptin in corneas from keratoconus and infectious keratitis patients. *J Ocular Pharmacol Ther* 2005;21(5):382-387.
- 17 Gariano RF, Nath AK, D'Amico DJ, Lee T, Sierra-Honigmann MR. Elevation of vitreous leptin in diabetic retinopathy and retinal detachment. *Invest Ophthalmol Vis Sci* 2000;41(11):3576-3581.
- 18 Maberley D, Cui JZ, Matsubara JA. Vitreous leptin levels in retinal disease. *Eye (Lond)* 2006;20(7):801.

- 19 Maurya R, Bhattacharya P, Ismail N, Dagur PK, Joshi AB, Razdan K, McCoy JP, Ascher J, Dey R, Nakhasi HL. Differential role of leptin as an immunomodulator in controlling visceral leishmaniasis in normal and leptin-deficient mice. *Am J Trop Med Hyg* 2016;95(1):109-119.
- 20 Suganami E, Takagi H, Ohashi H, Suzuma K, Suzuma I, Oh H, Watanabe D, Ojima T, Suganami T, Fujio Y, Nakao K, Ogawa Y, Yoshimura N. Leptin stimulates ischemia-induced retinal neovascularization: possible role of vascular endothelial growth factor expressed in retinal endothelial cells. *Diabetes* 2004;53(9):2443-2448.
- 21 Afarid M, Attarzadeh A, Farvardin M, Ashraf H. The association of serum leptin level and anthropometric measures with the severity of diabetic retinopathy in type 2 diabetes mellitus. *Med Hypothesis Discov Innov Ophthalmol* 2018;7(4):156-162.
- 22 Pult H, Wolffsohn JS. The development and evaluation of the new Ocular Surface Disease Index-6. *Ocular Surf* 2019;17(4):817-821.
- 23 van Bijsterveld OP. Diagnostic tests in the Sicca syndrome. *Arch Ophthalmol Chic Ill* 1969;82(1):10-14.
- 24 Oltulu P, Oltulu R, Asil M, Satirtav G, Mirza E. Conjunctival impression cytology and dry eye in patients with ulcerative colitis: a pilot study. *Eye Contact Lens* 2018;44 (Suppl 1):S190-S193.
- 25 Christen T, Trompet S, Noordam R, van Klinken JB, van Dijk KW, Lamb HJ, Cobbaert CM, den Heijer M, Jazet IM, Jukema JW, Rosendaal FR, de Mutsert R. Sex differences in body fat distribution are related to sex differences in serum leptin and adiponectin. *Peptides* 2018;107:25-31.
- 26 Frederich RC, Hamann A, Anderson S, Löllmann B, Lowell BB, Flier JS. Leptin levels reflect body lipid content in mice: evidence for diet-induced resistance to leptin action. *Nat Med* 1995;1(12):1311.
- 27 Margetic S, Gazzola C, Pegg G, Hill R. Leptin: a review of its peripheral actions and interactions. *Int J Obes* 2002;26(11):1407.
- 28 Ostlund RE, Yang JW, Klein S, Gingerich R. Relation between plasma leptin concentration and body fat, gender, diet, age, and metabolic covariates. *J Clin Endocrinol Metab* 1996;81(11):3909-3913.
- 29 Gröschl M, Wagner R, Dörr HG, Blum W, Rascher W, Dötsch J. Variability of leptin values measured from different sample matrices. *Horm Res* 2000;54(1):26-31.
- 30 Frank S, Stallmeyer B, Kämpfer H, Kolb N, Pfeilschifter J. Leptin enhances wound re-epithelialization and constitutes a direct function of leptin in skin repair. *J Clin Invest* 2000;106(4):501-509.
- 31 El-Deeb AM, Fansa HA, Wahba OM. The expression of leptin in oral wound healing in diabetes mellitus: an experimental study. *Int J Heal Sci* 2018;12(2):22.
- 32 Wu ZS, Shao P, Dass CR, Wei YZ. Systemic leptin administration alters callus VEGF levels and enhances bone fracture healing in wildtype and ob/ob mice. *Injury* 2018;49(10):1739-1745.
- 33 Lee M, Lee E, Jin SH, Ahn S, Kim SO, Kim J, Choi D, Lim KM, Lee ST, Noh M. Leptin regulates the pro-inflammatory response in human epidermal keratinocytes. *Arch Dermatol Res* 2018;310(4):351-362.